



Biodiversity evaluation with eDNA

Missions 2022 with Beauval Nature

Sabah – Bornéo in collaboration with the association of **HUTAN** and **Danau Girand Field Centre**.

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Preamble

This mission follows on from an initial test conducted in 2019 on the Kinabatangan River and its tributaries. This initial study enabled the eDNA method to be evaluated in conditions typical of Borneo's tropical climate, the results of different primers to be analyzed, the distribution of biodiversity along the river to be studied, and HUTAN teams to be trained in the use of this innovative method.

In 2022, two series of samples were taken from different areas studied by HUTAN, including oil palm plantations, ecological corridors, and forests less impacted by human activity. The vertebrate primer was used to verify the effectiveness of the eDNA method in biodiversity monitoring protocols. It was also used to evaluate the detection capacity of certain cryptic species compared to methods traditionally used to conduct wildlife inventories.

Additional data was collected from several aquatic environments, both from sites already surveyed in the past in order to analyze changes in results, and from new areas favorable to certain fish species in order to search for a discreet species, the kaloi.



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Project objectives

- Evaluate the effectiveness of eDNA in stagnant water (2nd campaign) compared to running water (1st campaign),
- Evaluate biodiversity at various treatment sites monitored by Hutan teams,
- Quickly collect data for comparison with data already acquired by Hutan as part of their long-term biodiversity monitoring,
- Detect rare and/or cryptic species,
- Train Hutan teams in the eDNA approach,
- Evaluate the feasibility of this new approach for biodiversity monitoring in an agricultural landscape dominated by oil palm plantations.



Figure 1: Palm oil plantation and forest

eDNA methodology

Protocol :

Thirty-two Spygen stagnant water ladle kits were used. We can analyze 400 meters of banks in a stagnant aquatic site with one kit (ponds, lakes, marshes, etc.).

Two liters of water are collected using a ladle, with 20 meters separating two samples (Spygen stagnant water protocol). The collected water is then filtered to collect eDNA. After filtration, a buffer solution preserved in a buffer solution until laboratory analysis in France.

Once at the laboratory, the eDNA from the filters is extracted and then amplified by PCR.

The laboratory then transmits the results for the different MOTUs identified. A MOTU (Molecular Operational Taxonomic Unit) is compared with sequences contained in the public reference database GenBank. In

some cases, the comparison allows us to identify the MOTU down to the species level, but in others, the MOTU is identified at lower levels: *Genus, Family, Order, Class*, etc.

Here, we have kept only the MOTUs with the Species and Genus levels. A review of the results and a comparison with existing knowledge of species distribution allowed us to refine some of the MOTUs.

We opted for this stagnant water kit to analyze several environments, including lakes, marshes, ditches, and puddles. Indeed, depending on the habitats examined, the water may be drained, collected in ditches such as in oil palm plantations, or, quite rarely, found as puddles in forests with little human impact.



Figure 2 : Mangrove in a brackish environment



Figure 3 : Oxbow lake

Sampling plan



Sampling was carried out in different landscapes, all of which were studied by Hutan:

- Industrial palm plantations,
- Agroforestry landscapes (trees and palms planted together at the same time),
- Restored ecological corridors between isolated forest blocks at different stages of regeneration,
- Oxbow lakes,
- Natural forests,

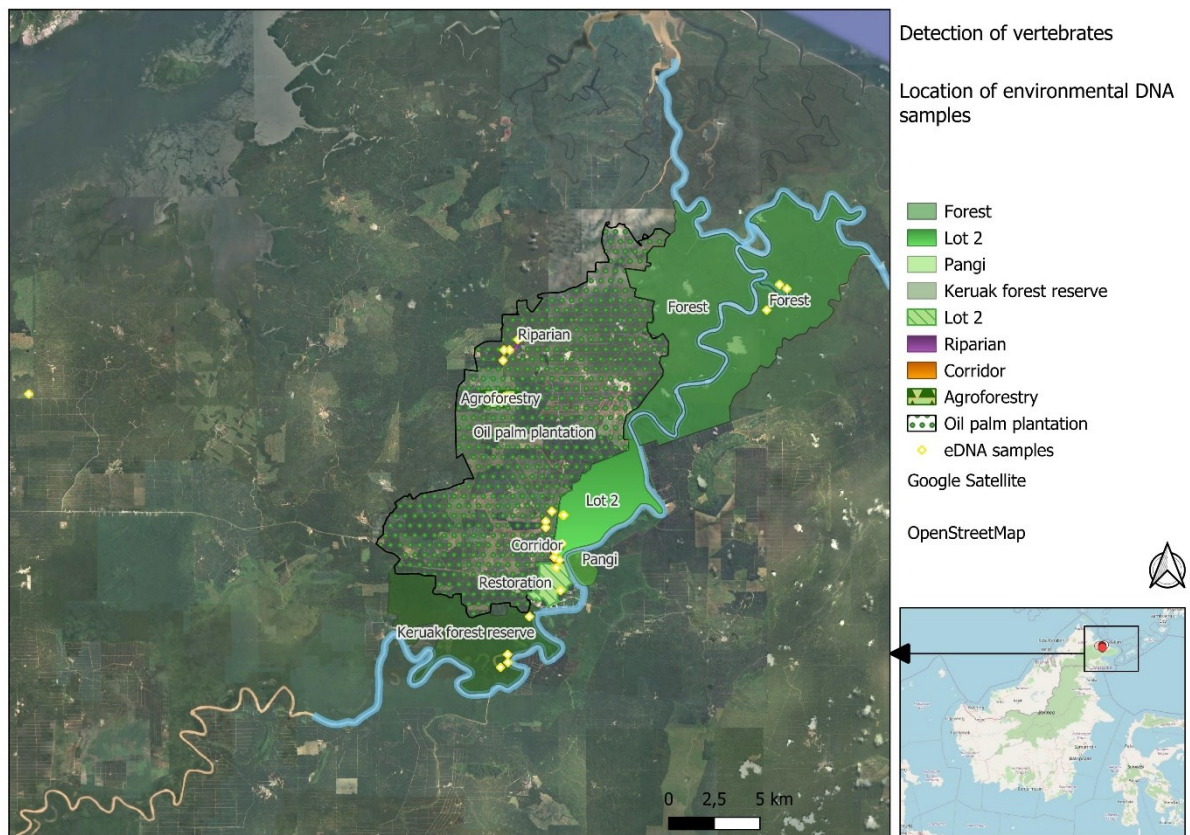


Figure 4 : Location of eDNA sampling sites

Overall results

There are several levels to read the results

This section provides an overview of the results obtained. In total, samples of the second campaign identified 84 species, with mammals representing the most diverse class with 33 different species, followed by birds (25), fish (13), amphibians (9), and reptiles (4): Figure 5. Unlike fish, mammals are generally more studied and better identified in genetic databases, contributing to a significantly higher identification rate.

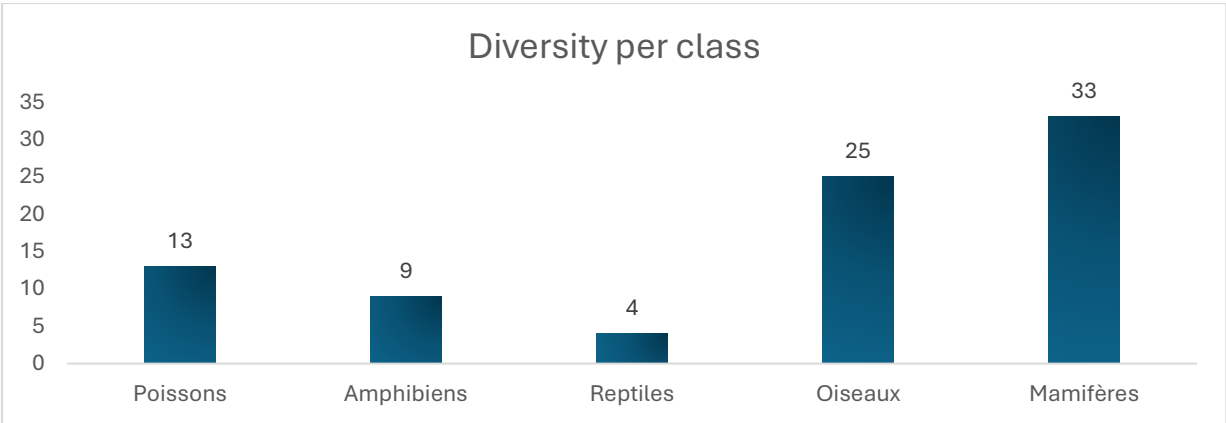


Figure 5 : Diversity per class : Fish, Amphibians, Reptiles, Birds, Mammals

In terms of the IUCN vulnerability risks, **25% of the species are vulnerable to critically endangered**: Fig. 6.

IUCN vulnerability level

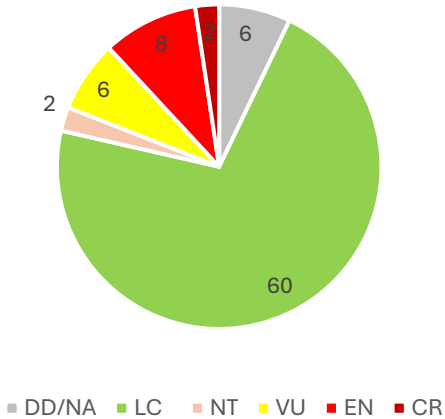


Figure 6 : Levels of vulnerability of species detected

Species with a conservation status of Vulnerable (VU) or higher are specially mapped and an extract from their IUCN data sheet is provided below.

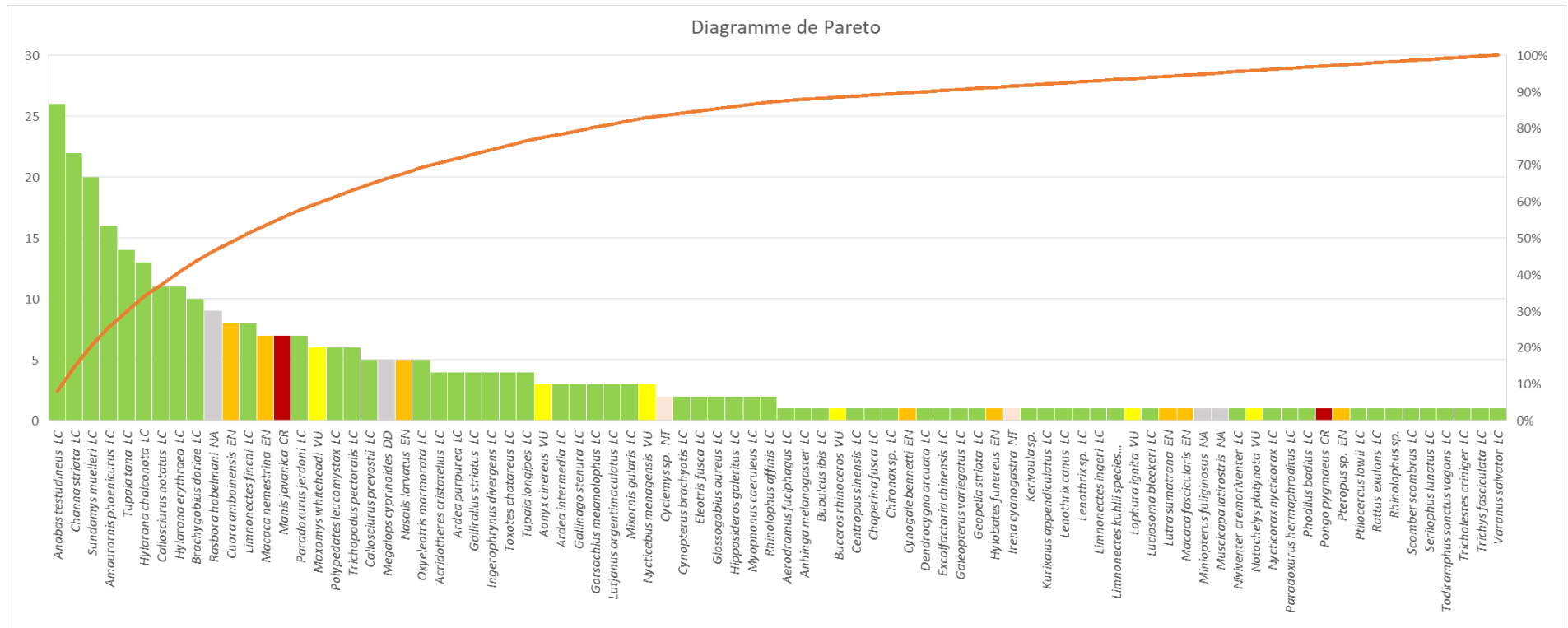


Figure 7 : Pareto chart

The Pareto chart shows the frequency of species detection. It allows to visualize and evaluate the representativeness of each species across all samples collected. For example, the graph indicates that the Mueller's rat, *Sundamys muelleri* is detected in 20% of the samples.



The terrestrial species that are the most frequently identified in the Pareto chart (score >10% or even >5%) are all common species regularly detected by Hutan teams during biodiversity monitoring. These species are generalists and highly adaptable, explaining their relative abundance in eDNA inventories.

- **Fish** : the two common fish species are identical to the results from our 2019 analyses. **Anabas testudineus (LC)**, the climbing perch, is present in over 25% of our surveys. It is a freshwater fish species adapted to low-oxygen environments. Individuals of this species are very resilient and can even survive out of water for several days and move across land between two bodies of water. This fish is therefore very common and is found in all the environments tested. Another fish species that is very common in our samples is **Channa striata (LC)** or Snakehead murrel (Haruan). This carnivorous species is widespread throughout most of Asia. This top predator can also survive in muddy conditions because it breathes air through its skin. It is interesting to note that the Bumblebee Goby (*Brachydorius doriae*) is also commonly found in our samples. This small fish is prized by aquarium enthusiasts because of its yellow and black colouring. It can be found in clear, turbid and polluted drain water.

- **Mammals** : small mammals species like the Muller's rat, the common tree shrew and the plantain squirrel are common species that Hutan teams have identified in all environments included in the biodiversity monitoring plots. This abundance is reflected in the results of the Pareto diagram, as these species are present in more than 10% of the sampled sites.
- **Amphibians** : the two species classed in the top group (>10%) are also common species found particularly in degraded and heavily anthropized areas.
- **Birds** : one bird species, the white-bellied waterhen was prevalent in our eDNA samples. This species is dependent on open degraded aquatic environments, which explains its abundance in our results.



The commonest endangered species in our samples is a turtle, ***Cuora amboinensis (EN)***, or Southeast Asian Box Turtle. This species is common in the Kinabatangan but is declining in many areas due to trafficking for meat and captivity.

The Pig-tailed macaque, very common in the Kinabatangan, moves across very large areas, explaining its high score on the Pareto chart (figure 7). This species of macaques, which is endangered in many areas of its range, is quite common in kinabatangan. It moves mainly on the ground, explaining the abundance of genetic material collected in our samples.

The Venn diagram identifies the specific and common diversity of the three environments studied: the forest, the oil palm plantation and the corridor created to connect two forest island.

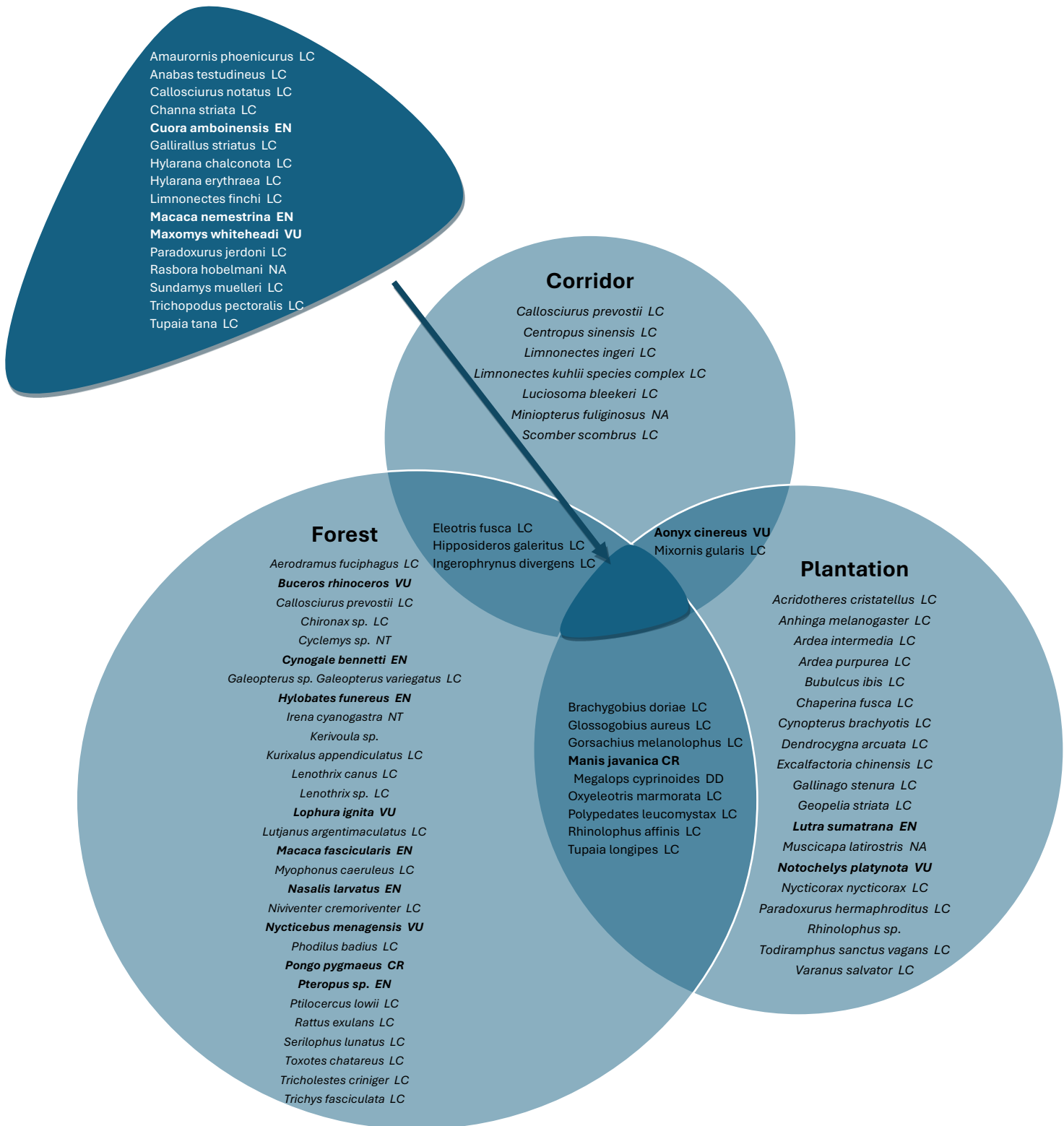


Figure 8 : Venn diagram

As expected, species found at the intersection of the three habitat types are mostly generalists that can survive and adapt fairly well to habitat disturbance. It is noteworthy that we detected the endangered *Lutra sumatrana* only in the plantations, while the CE pangolin was found not only in plantations but also in the forest (see below).

Figure 8 : List of taxa identified down to the species level

Classes	Ordres	Familles	Espèces
Actinopteri	Anabantiformes	Anabantidae	Anabas testudineus LC
		Channidae	Channa striata LC
		Osphronemidae	Trichopodus pectoralis LC
	Cypriniformes	Danionidae	Luciosoma bleekeri LC
			Rasbora hobelmani NA
	Elopiformes	Megalopidae	Megalops cyprinoides DD
		Eleotridae	Eleotris fusca LC
	Gobiiformes		Gobiidae
		Brachygobius doriae LC	
		Glossogobius aureus LC	
	Lutjaniformes	Lutjanidae	Lutjanus argentimaculatus LC
	Scombriformes	Scombridae	Scomber scombrus LC
(vide)	Toxotidae	Toxotes chatareus LC	
Aves	Anseriformes	Anatidae	Dendrocygna arcuata LC
	Apodiformes	Apodidae	Aerodramus fuciphagus LC
	Bucerotiformes	Bucerotidae	Buceros rhinoceros VU
	Charadriiformes	Scolopacidae	Gallinago stenura LC
			Geopelia striata LC
	Coraciiformes	Alcedinidae	Todiramphus sanctus vagans LC
	Cuculiformes	Centropidae	Centropus sinensis LC
	Galliformes	Phasianidae	Excalfactoria chinensis LC
			Lophura ignita VU
		Gruiformes	Rallidae
	Gallirallus striatus LC		

	Passeriformes	Eurylaimidae	Serilophus lunatus LC
		Irenidae	Irena cyanogastra NT
		Muscicapidae	Muscicapa latirostris NA
		Myophonus caeruleus LC	
		Pycnonotidae	Tricholestes criniger LC
		Sturnidae	Acridotheres cristatellus LC
	Pelecaniformes	Timaliidae	Mixornis gularis LC
		Anhingidae	Anhinga melanogaster LC
		Ardeidae	Ardea intermedia LC
		Ardeidae	Ardea purpurea LC
		Ardeidae	Bubulcus ibis LC
		Ardeidae	Gorsachius melanolophus LC
	Strigiformes	Ardeidae	Nycticorax nycticorax LC
Tytonidae		Phodilus badius LC	
Amphibia	Anura	Bufo	Ingerophrynus divergens LC
		Dicroglossidae	Limnonectes finchi LC
			Limnonectes ingeri LC
			Limnonectes kuhlii species complex LC
		Microhylidae	Chaperina fusca LC
		Ranidae	Hylarana chalconota LC
			Hylarana erythraea LC
		Rhacophoridae	Kurixalus appendiculatus LC
Polypedates leucomystax LC			
Mammalia	Carnivora	Mustelidae	Aonyx cinereus VU
		Lutra sumatrana EN	
		Cynogale bennetti EN	
		Viverridae	Paradoxurus jerdoni LC
	Paradoxurus hermaphroditus LC		
	Chiroptera	Hipposideridae	Hipposideros galeritus LC
		Chironax sp. LC	
		Pteropodidae	Cynopterus brachyotis LC

			Pteropus sp. EN
		Rhinolophidae	Rhinolophus affinis LC Rhinolophus sp.
		Vespertilionidae	Kerivoula sp. Miniopterus fuliginosus NA
Dermoptera	Cynocephalidae		Galeopterus variegatus LC
Pholidota	Manidae		Manis javanica CR
			Macaca fascicularis EN
	Cercopithecidae		Macaca nemestrina EN
Primates			Nasalis larvatus EN
	Hominidae		Pongo pygmaeus CR
	Hylobatidae		Hylobates funereus EN
	Lorisidae		Nycticebus menagensis VU
		Hystricidae	Trichys fasciculata LC
			Lenothrix canus LC
			Lenothrix sp. LC
			Maxomys whiteheadi VU
Rodentia	Muridae		Niviventer cremoriventer LC
			Rattus exulans LC
			Sundamys muelleri LC
		Sciuridae	Callosciurus notatus LC
			Callosciurus prevostii LC
			Ptilocercus lowii LC
Scandentia	Tupaiaidae		Tupaia longipes LC
			Tupaia tana LC
	Squamata	Varanidae	Varanus salvator LC
Reptilia			Cuora amboinensis EN
	Testudines	Geoemydidae	Cyclemys sp. NT
			Notochelys platynota VU

When reading the list of species detected, it appears that some species are common to all three habitats studied, some of which have a proven level of vulnerability, such as *Cuora amboinensis* (EN), *Macaca nemestrina* (EN) and *Maxomys whiteheadi* (VU). All these species are characterised by proven properties of adaptation to environmental degradation; they are mainly terrestrial but live close to wetlands.

It is surprising to note that common species that are regularly detected by our teams in all the environments studied have only been identified in one or two environments using eDNA. For example :

- ***Ardea intermedia*** (middle egret), ***Bulbulus ibicus*** (cattle egret), ***Varanus salvator*** (monitor lizard): only found in plantations, whereas these species, which are mainly aquatic, are present in all ecosystems in Kinabatangan
- ***Macaca fascicularis*** (long-tailed macaque) is only found in forests, although it is present in all environments.

This finding is explained by the fact that eDNA method is more effective with mammals and fish rather than reptiles and birds, as the former species release more genetic material into the environments than the later ones.

Several remarkable results:

- ***Lutra sumatrana*** (hairy nosed otter): this extremely elusive species was only detected in oil palm plantations. This finding is noteworthy since only a few records of this species exist for Kinabatangan and all in forest or oxbow lakes.
- ***Scomber scombrus*** (mackerel): this species is widely consumed by people in canned food or fresh (from the market) and appear thus to be a “pollutant” from food-related origin,
- **Orang-outang**: the species was only detected in the forest, although it also ranges in oil palm plantations and forest corridors.
- Surprisingly, we failed to detect any DNA from the Asian elephant, a common and generalist species found in all habitats in Kinabatangan. However, this species is currently absent from the genetic database used for this study, explaining the result. For this species, eDNA is not useful to study this species, is not a cryptic species.

Unsurprisingly, we identified more species in the forest, this environment being the richest in endangered or vulnerable species.

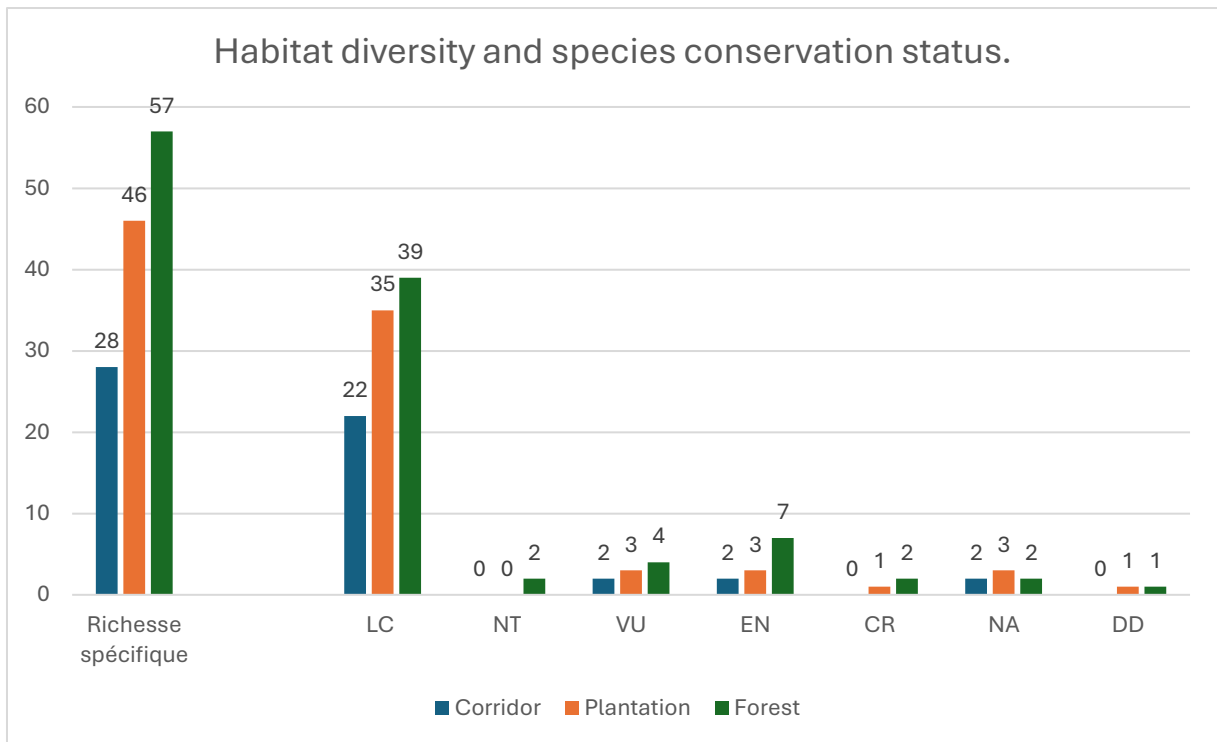


Figure 9 : Habitat diversity and species conservation status.

The figure above (Fig. 9) shows another interpretation of the results. When comparing the three habitats studied, the forest shows the most diverse and a greater number of animals species with an unfavourable conservation status.

The eDNA results concerning the ecological corridor contrast a lot with the results of the biodiversity monitoring carried out by Hutan teams. First, the latter indicates a number of intermediate species between forest and plantations, whereas the eDNA result indicates a lower number of species in the corridor. It's important to remember that the role of the corridor is not to accommodate species so they can complete their entire life cycle, but rather to provide passageways between forest islands. Thus, the presence of animal species on the corridor is therefore short-lived. They are merely passing through and, as a result, leave proportionally less genetic material behind, which may explain this low eDNA diversity. For this reason, the eDNA methodology may not be the most suitable tool for verifying the effectiveness of the corridor for wildlife.

The graph (figure 10) below models the differences between the three habitats with a projection of the results (100 samples).

As expected, the forest habitat has the greatest diversity and variability (very poor Forest 2 and very high diversity Forest1 and Forest lot 2).

Old plantation (palm old) appears to be less diverse than the forest (Forest 1 and Forest lot 2) but more diverse than young plantations. Indeed, the tridimensional environment characterizing mature plantations can accommodate a certain number of species that are not found in the early

development stages of an oil palm plantation. Finally, corridors, as shown by the results of Hutan's biodiversity monitoring, appear to constitute an intermediate habitat between forest and plantation.

Rarefaction / Extrapolation Curves Species richness per plot

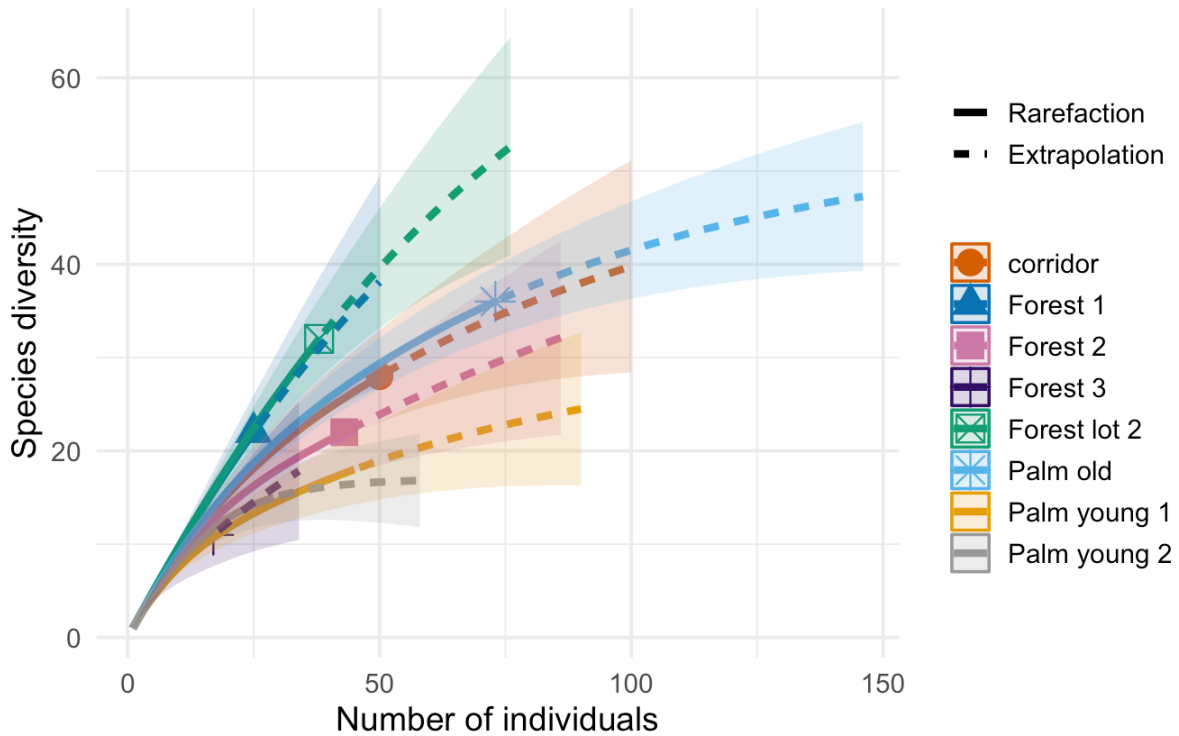


Figure 10 : Extrapolation curves. Richness per habitat based on the number of samples.

After assessing habitat diversity and its potential role in the conservation of endangered species, we will look at the proportion of animal classes, and their diets present in each habitat.

Animal composition of the three habitats studied

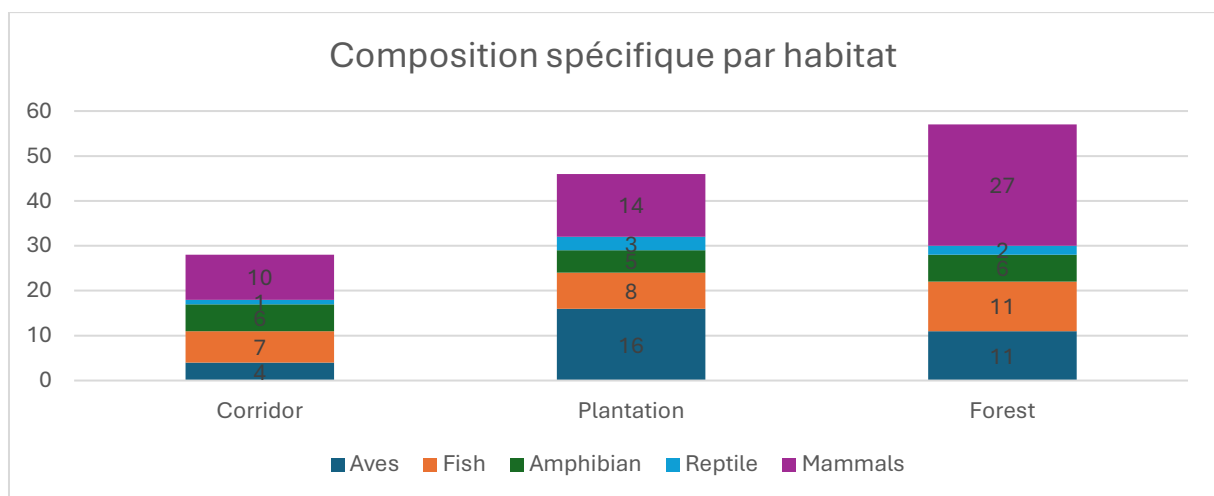


Figure 11 : Specific composition by habitat

We will systematically compare the results of the plantations and the corridor with those obtained in the **forest environment**, which is the **reference** state for biodiversity in Kinabatangan.

Even degraded forests play an important role in the survival of certain animal species that cannot survive in non-forest environments.

A more detailed functional analysis would enable species assemblages to be differentiated according to the lifestyles of the species detected (arboreal, burrowing, flying, etc.), body mass or diet.

For the moment, only the **diet** of the species identified in each habitat will be presented and compared, as well as the faunal composition by class.



FOREST

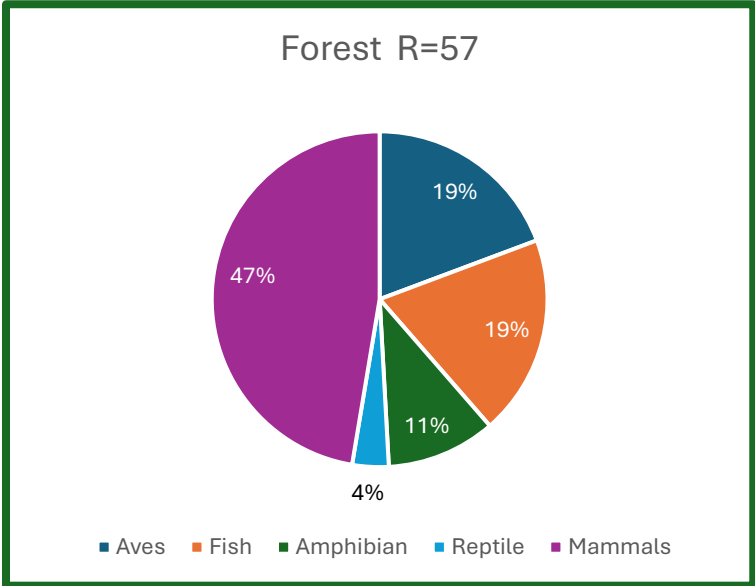


Figure 12 : Fauna composition of species detected in the forest class.

Nearly 50% of the species detected in the forest habitat are mammals. Birds and fish are half as common, followed by amphibians and finally reptiles.

These results are explained by the recent presence of the species in this ecosystem and their ability to disseminate genetic material in their environment.

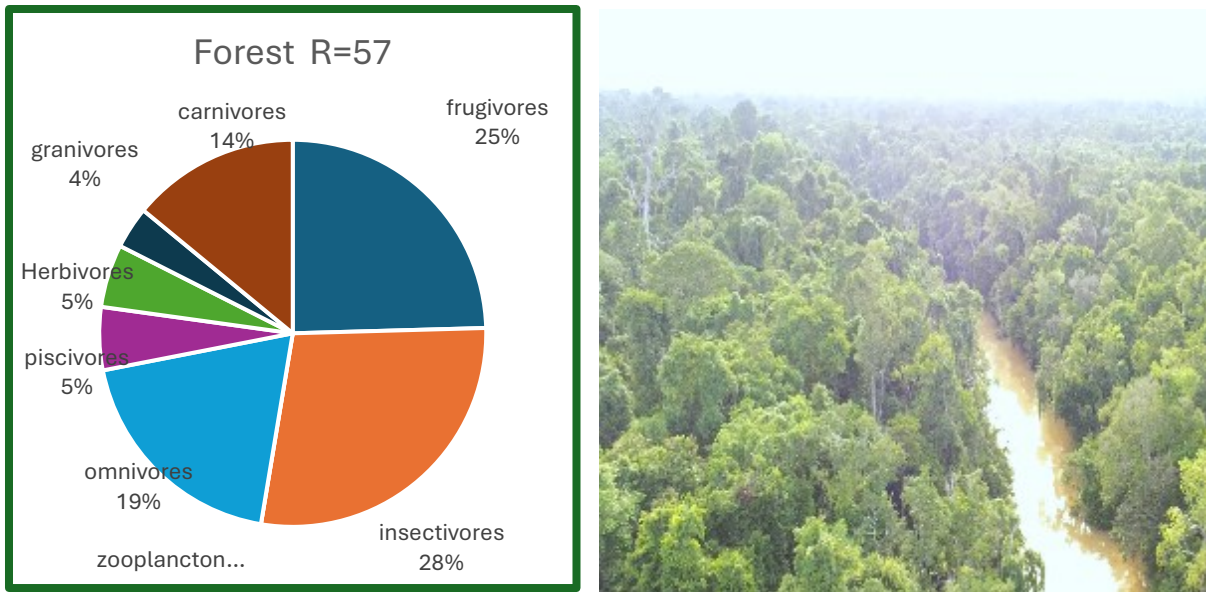


Figure 13 : Fauna composition of species detected in forests by diet

The diets of animals detected in the forest are divided between frugivores and insectivores, omnivores and carnivores.

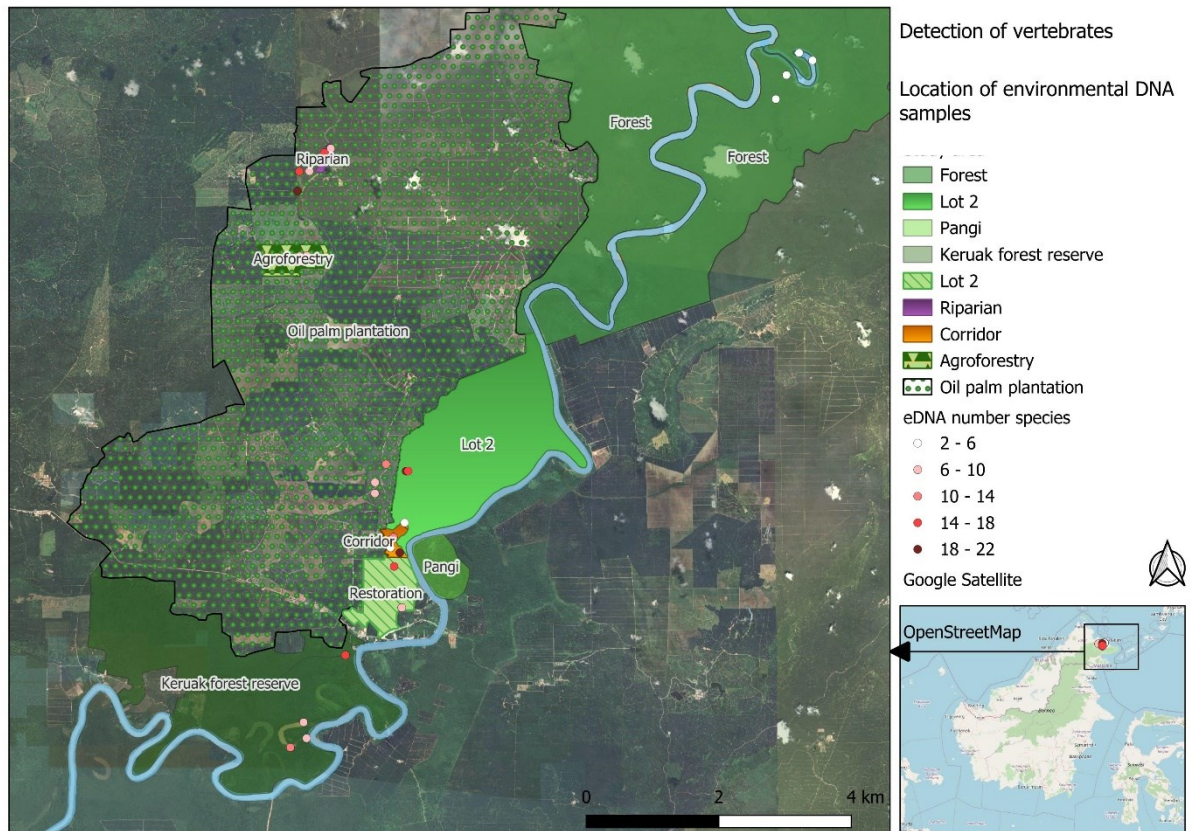


Figure 14 : Location map of analysed forests

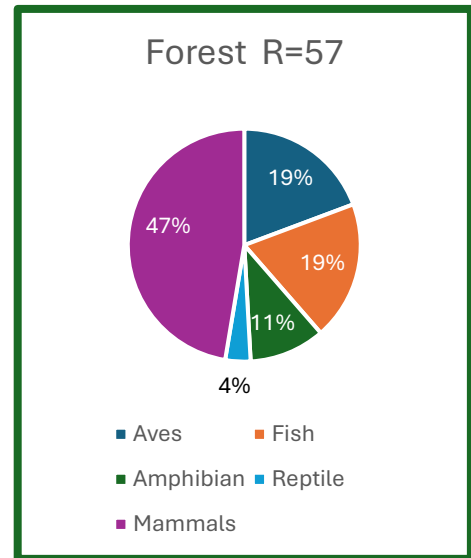
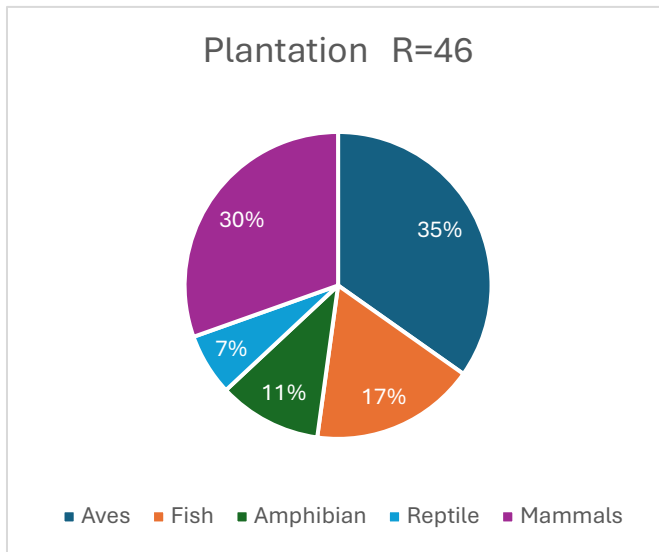
From South to North, samples were taken in the Keruak forest reserve, then in an area undergoing restoration, in Lot 2 of the Lower Kinabatangan Wildlife Sanctuary and further north on an oxbow lake surrounded by a relatively undisturbed forest.

Species	Class	Diet
<i>Aerodramus fuciphagus</i> LC	Aves	insectivores
<i>Buceros rhinoceros</i> VU	Aves	frugivores
<i>Callosciurus prevostii</i> LC	Mammalia	frugivores
<i>Chironax</i> sp. LC	Mammalia	frugivores
<i>Cyclemys</i> sp. NT	Reptilia	carnivores
<i>Cynogale bennetti</i> EN	Mammalia	piscivores
<i>Galeopterus</i> sp. <i>Galeopterus variegatus</i> LC	Mammalia	Herbivores
<i>Hylobates funereus</i> EN	Mammalia	omnivores
<i>Irena cyanogastra</i> NT	Aves	frugivores
<i>Kerivoula</i> sp.	Mammalia	insectivores
<i>Kurixalus appendiculatus</i> LC	Amphibia	insectivores
<i>Lenothrix canus</i> LC	Mammalia	frugivores
<i>Lenothrix</i> sp. LC	Mammalia	frugivores
<i>Lophura ignita</i> VU	Aves	insectivores
<i>Lutjanus argentimaculatus</i> LC	Actinopteri	piscivores
<i>Macaca fascicularis</i> EN	Mammalia	frugivores
<i>Myophonus caeruleus</i> LC	Aves	insectivores
<i>Nasalis larvatus</i> EN	Mammalia	frugivores
<i>Niviventer cremoriventer</i> LC	Mammalia	granivores
<i>Nycticebus menagensis</i> VU	Mammalia	omnivores
<i>Phodilus badius</i> LC	Aves	carnivores
<i>Pongo pygmaeus</i> CR	Mammalia	frugivores
<i>Pteropus</i> sp. EN	Mammalia	frugivores
<i>Ptilocercus lowii</i> LC	Mammalia	frugivores
<i>Rattus exulans</i> LC	Mammalia	omnivores
<i>Serilophus lunatus</i> LC	Aves	insectivores
<i>Toxotes chatareus</i> LC	Actinopteri	carnivores
<i>Tricholestes criniger</i> LC	Aves	granivores
<i>Trichys fasciculata</i> LC	Mammalia	omnivores
<i>Eleotris fusca</i> LC	Actinopteri	carnivores
<i>Hipposideros galeritus</i> LC	Mammalia	insectivores
<i>Ingerophrynus divergens</i> LC	Amphibia	insectivores
<i>Brachygobius doriae</i> LC	Actinopteri	carnivores
<i>Glossogobius aureus</i> LC	Actinopteri	carnivores
<i>Gorsachius melanolophus</i> LC	Aves	insectivores
<i>Megalops cyprinoides</i> DD	Actinopteri	piscivores
<i>Oxyeleotris marmorata</i> LC	Actinopteri	omnivores
<i>Polypedates leucomystax</i> LC	Amphibia	insectivores
<i>Rhinolophus affinis</i> LC	Mammalia	insectivores

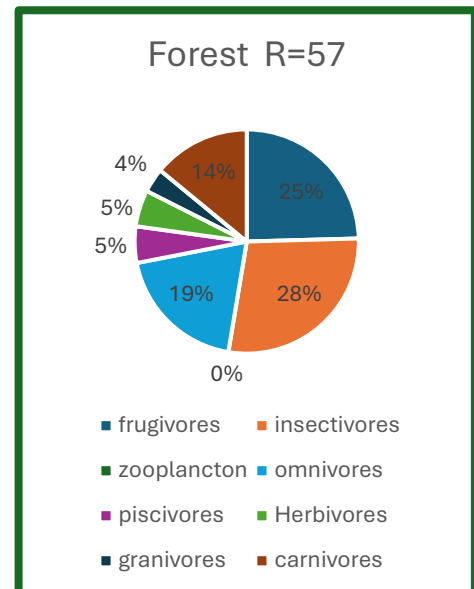
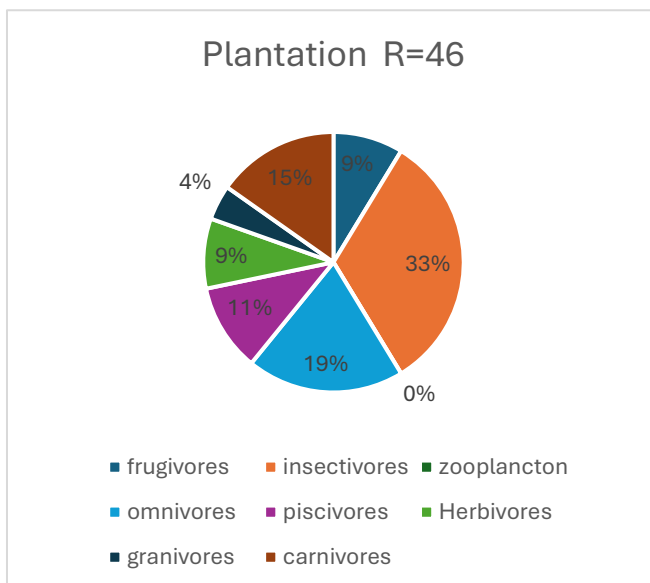
<i>Tupaia longipes</i> LC	Mammalia	omnivores
<i>Manis javanica</i> CR	Mammalia	insectivores
<i>Amaurornis phoenicurus</i> LC	Aves	insectivores
<i>Anabas testudineus</i> LC	Actinopteri	Herbivores
<i>Callosciurus notatus</i> LC	Mammalia	frugivores
<i>Channa striata</i> LC	Actinopteri	carnivores
<i>Cuora amboinensis</i> EN	Reptilia	Herbivores
<i>Gallirallus striatus</i> LC	Aves	carnivores
<i>Hylarana chalconota</i> LC	Amphibia	insectivores
<i>Hylarana erythraea</i> LC	Amphibia	insectivores
<i>Limnonectes finchi</i> LC	Amphibia	insectivores
<i>Macaca nemestrina</i> EN	Mammalia	frugivores
<i>Maxomys whiteheadi</i> VU	Mammalia	omnivores
<i>Paradoxurus jerdoni</i> LC	Mammalia	frugivores
<i>Rasbora hobelmani</i> NA	Actinopteri	omnivores
<i>Sundamys muelleri</i> LC	Mammalia	omnivores
<i>Trichopodus pectoralis</i> LC	Actinopteri	omnivores
<i>Tupaia tana</i> LC	Mammalia	omnivores

Figure 15 : List of species detected in the forest

PLANTATION



Plantations are seeing a decline in their mammal diversity, with birds becoming more prevalent. It is hypothesised that the decline in mammals in oil palm plantations is due to drier conditions (Kay and Madden 1997); lack of suitable resting or feeding resources; competition with domestic animals; hunting and killing; etc.



The proportion of **frugivores** falls from 25% in the forest to 9% in plantations, showing the rarefaction of flower diversity in agricultural landscapes. In the contrary the insectivore proportion tends to increase very slightly.

For frugivores, the result can be explained by the presence of only one type of fruit tree in plantations. Besides the fruits of oil palm, there is relatively little other fruit available. In tropical forest in the contrary, frugivores have access to a significantly higher diversity of fruits than in a monoculture habitat ([Kissling, Gaese, 2009](#)).

We also note a decline (although less severe than for frugivorous) of insectivore species, in plantations compared to the forests.

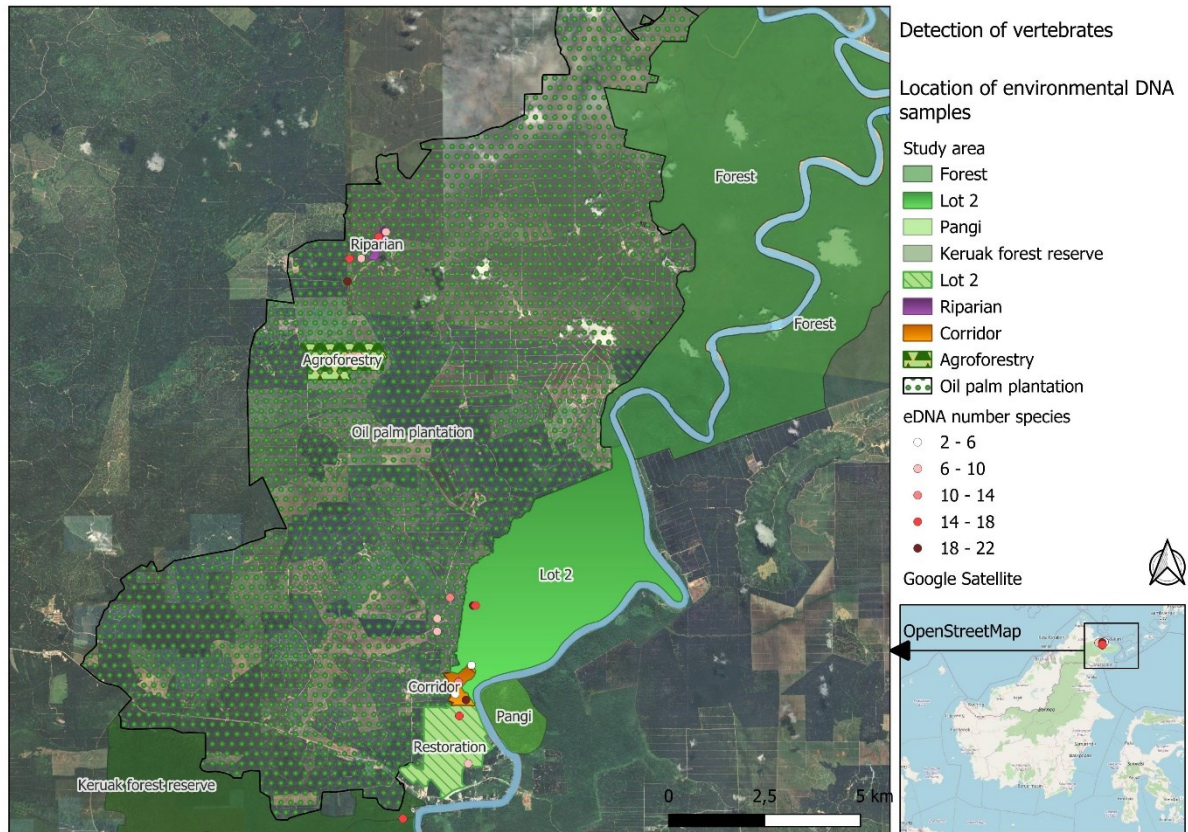
Indeed, former studies have already shown a decline in insectivore diversity in the palm oil plantation. Insectivore abundance and species richness are greater in degraded forest than in palm oil plantations (Soon et al, 2022), stressing out the importance of maintaining natural forests within and around plantations to increase the diversity of insectivores in plantation.



Figure 16 : Aerial view of a single-species oil palm plantation



Figure 17 : Plantation with forest island



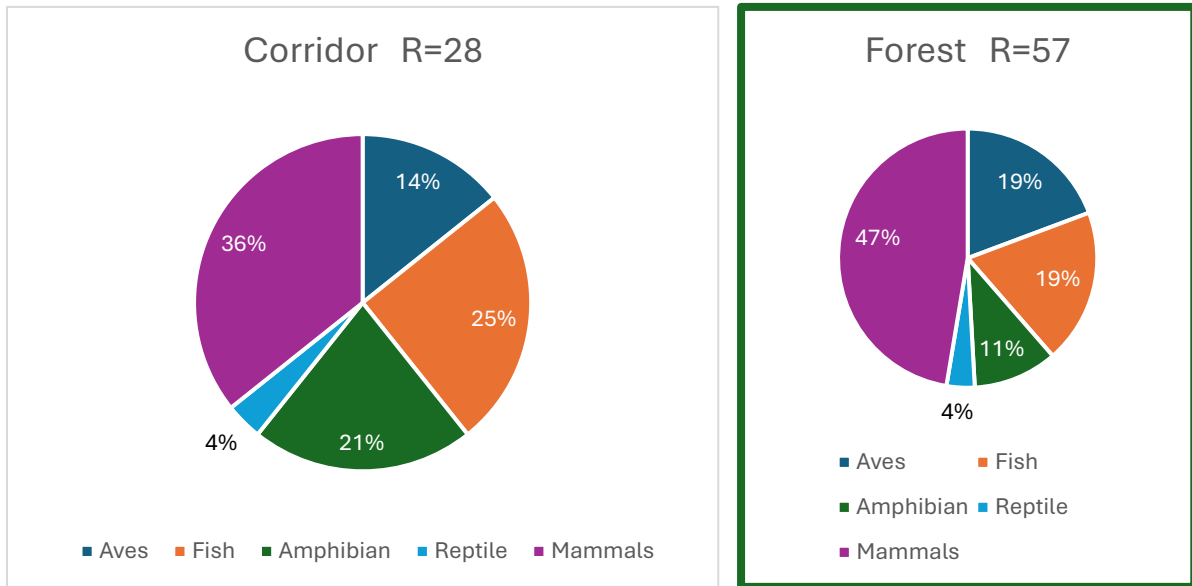
Samples were taken from different plots of oil palm trees of varying ages.

Species	Class	Diet
<i>Acridotheres cristatellus</i> LC	Aves	omnivores
<i>Anhinga melanogaster</i> LC	Aves	piscivores
<i>Ardea intermedia</i> LC	Aves	piscivores
<i>Ardea purpurea</i> LC	Aves	piscivores
<i>Bubulcus ibis</i> LC	Aves	insectivores
<i>Chaperina fusca</i> LC	Amphibia	insectivores
<i>Cynopterus brachyotis</i> LC	Mammalia	frugivores
<i>Dendrocygna arcuata</i> LC	Aves	Herbivores
<i>Excalfactoria chinensis</i> LC	Aves	granivores
<i>Gallinago stenura</i> LC	Aves	insectivores
<i>Geopelia striata</i> LC	Aves	granivores
<i>Lutra sumatrana</i> EN	Mammalia	carnivores
<i>Muscicapa latirostris</i> NA	Aves	insectivores
<i>Notochelys platynota</i> VU	Reptilia	Herbivores
<i>Nycticorax nycticorax</i> LC	Aves	piscivores
<i>Paradoxurus hermaphroditus</i> LC	Mammalia	carnivores
<i>Rhinolophus</i> sp.	Mammalia	insectivores
<i>Todiramphus sanctus vagans</i> LC	Aves	insectivores
<i>Varanus salvator</i> LC	Reptilia	omnivores
<i>Aonyx cinereus</i> VU	Mammalia	carnivores

<i>Mixornis gularis</i> LC	Aves	insectivores
<i>Brachygobius doriae</i> LC	Actinopteri	carnivores
<i>Glossogobius aureus</i> LC	Actinopteri	carnivores
<i>Gorsachius melanolophus</i> LC	Aves	insectivores
<i>Megalops cyprinoides</i> DD	Actinopteri	piscivores
<i>Oxyeleotris marmorata</i> LC	Actinopteri	omnivores
<i>Polypedates leucomystax</i> LC	Amphibia	insectivores
<i>Rhinolophus affinis</i> LC	Mammalia	insectivores
<i>Tupaia longipes</i> LC	Mammalia	omnivores
<i>Manis javanica</i> CR	Mammalia	insectivores
<i>Amaurornis phoenicurus</i> LC	Aves	insectivores
<i>Anabas testudineus</i> LC	Actinopteri	Herbivores
<i>Callosciurus notatus</i> LC	Mammalia	frugivores
<i>Channa striata</i> LC	Actinopteri	carnivores
<i>Cuora amboinensis</i> EN	Reptilia	Herbivores
<i>Gallirallus striatus</i> LC	Aves	carnivores
<i>Hylarana chalconota</i> LC	Amphibia	insectivores
<i>Hylarana erythraea</i> LC	Amphibia	insectivores
<i>Limnonectes finchi</i> LC	Amphibia	insectivores
<i>Macaca nemestrina</i> EN	Mammalia	frugivores
<i>Maxomys whiteheadi</i> VU	Mammalia	omnivores
<i>Paradoxurus jerdoni</i> LC	Mammalia	frugivores
<i>Rasbora hobelmani</i> NA	Actinopteri	omnivores
<i>Sundamys muelleri</i> LC	Mammalia	omnivores
<i>Trichopodus pectoralis</i> LC	Actinopteri	omnivores
<i>Tupaia tana</i> LC	Mammalia	omnivores

Figure 18 : List of species found in the plantations

CORRIDOR



We would like to point out that in this study, only one corridor with a limited surface could be analysed, limiting the conclusions of the analysis. .

About half as much species diversity was documented in the corridor compared to the forest, with a higher proportion of amphibians and fish than mammals. It should be noted that the corridor is an old plantation that was not cut down when it reached a stage of low or no productivity. However, this plot has been enriched with local plant to increase its tree diversity.

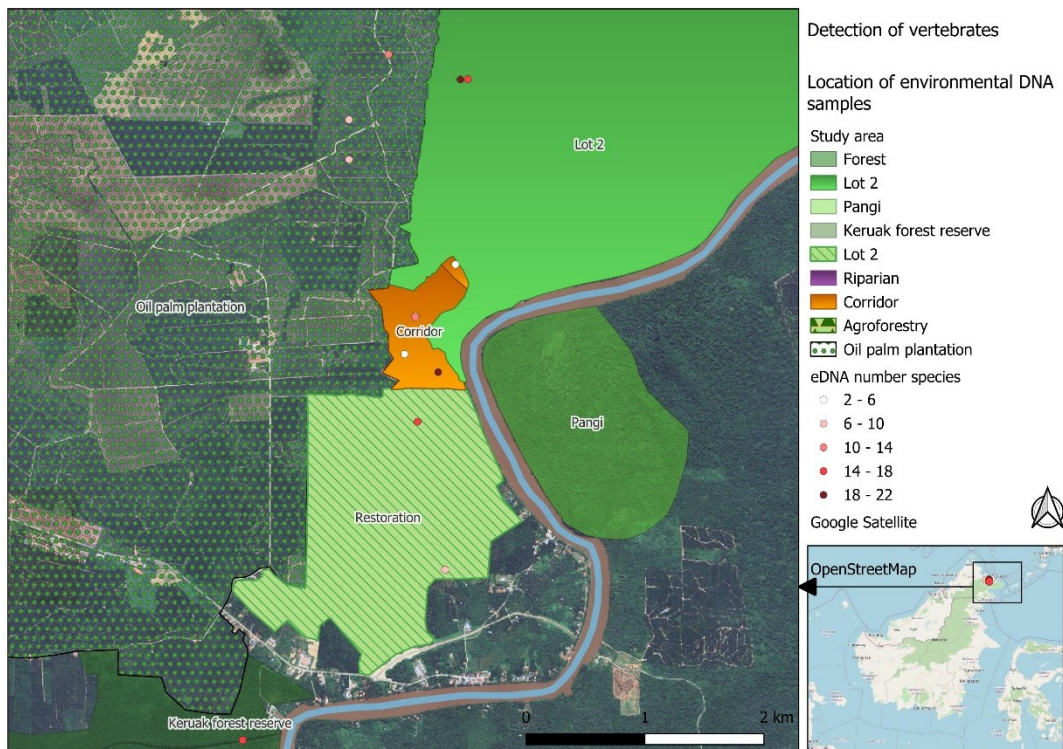
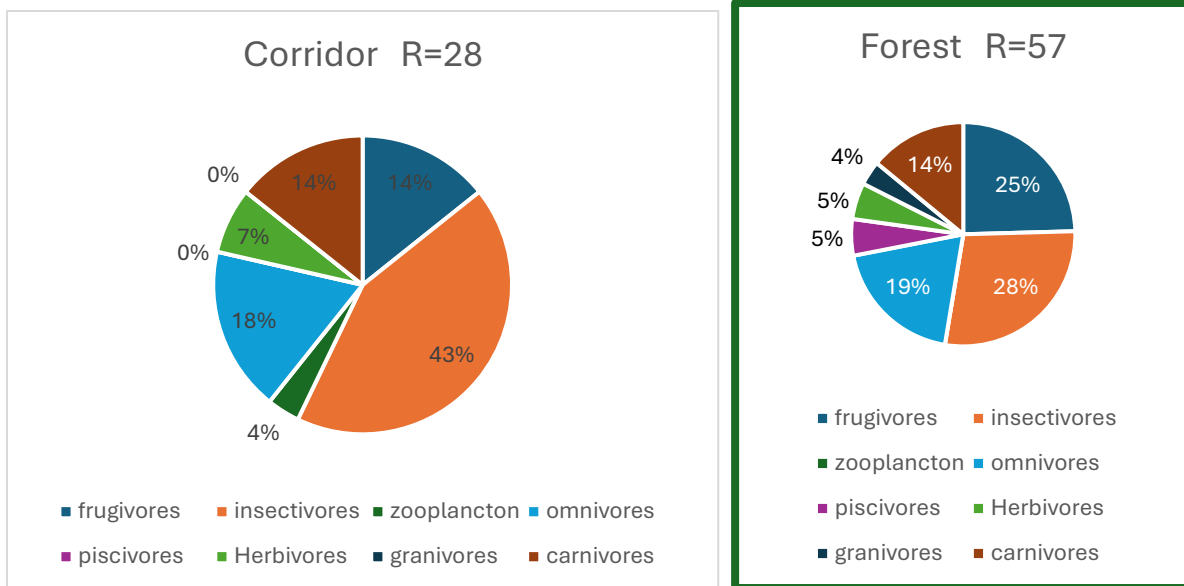


Figure 19 : Location of the forest corridor



This habitat has half the species diversity of the forest, but its insectivores proportion remains significant. We also detected several species of amphibians and bats that were not found anywhere else in this study.



Figure 20 :Example of a corridor created in a plantation after harvesting.

Here, the riverbanks are still lined with old palm trees that should have been cut down like those on neighbouring plot. This prevents the banks from being destabilised and also preserves a significant amount of vegetation along the watercourse.





Figure 21 : The isolated tree after cutting down a plot of cultivated palm trees.

Here, after palm tree harvesting, only one local tree has been preserved. Plantations around it would help restore area.

Species	Class	Diet
<i>Callosciurus prevostii</i> LC	Mammalia	frugivores
<i>Centropus sinensis</i> LC	Aves	insectivores
<i>Limnonectes ingeri</i> LC	Amphibia	insectivores
<i>Limnonectes kuhlii</i> species complex LC	Amphibia	insectivores
<i>Luciosoma bleekeri</i> LC	Actinopteri	insectivores
<i>Miniopterus fuliginosus</i> NA	Mammalia	insectivores
<i>Scomber scombrus</i> LC	Actinopteri	zooplankton
<i>Aonyx cinereus</i> VU	Mammalia	carnivores
<i>Mixornis gularis</i> LC	Aves	insectivores
<i>Eleotris fusca</i> LC	Actinopteri	carnivores
<i>Hipposideros galeritus</i> LC	Mammalia	insectivores
<i>Ingerophrynus divergens</i> LC	Amphibia	insectivores
<i>Amaurornis phoenicurus</i> LC	Aves	insectivores
<i>Anabas testudineus</i> LC	Actinopteri	Herbivores
<i>Callosciurus notatus</i> LC	Mammalia	frugivores
<i>Channa striata</i> LC	Actinopteri	carnivores
<i>Cuora amboinensis</i> EN	Reptilia	Herbivores
<i>Gallirallus striatus</i> LC	Aves	carnivores
<i>Hylarana chalconota</i> LC	Amphibia	insectivores
<i>Hylarana erythraea</i> LC	Amphibia	insectivores
<i>Limnonectes finchi</i> LC	Amphibia	insectivores
<i>Macaca nemestrina</i> EN	Mammalia	frugivores
<i>Maxomys whiteheadi</i> VU	Mammalia	omnivores
<i>Paradoxurus jerdoni</i> LC	Mammalia	frugivores
<i>Rasbora hobelmani</i> NA	Actinopteri	omnivores
<i>Sundamys muelleri</i> LC	Mammalia	omnivores
<i>Trichopodus pectoralis</i> LC	Actinopteri	omnivores
<i>Tupaia tana</i> LC	Mammalia	omnivores

Figure 22 : List of species detected in the corridor

IUCN technical data sheets for detected species with a confirmed vulnerability level.

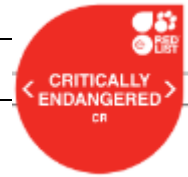
The focus is on the species with a confirmed vulnerability level. Mammals rank highest in the classification of vulnerable species. This class is the most studied and the most impacted by deforestation and habitat fragmentation.

Class	IUCN	Latin name - IUCN	
Mammalia	Critically endangered	<i>Manis javanica</i> CR	
Mammalia		<i>Pongo pygmaeus</i> CR	
Mammalia	Endangered	<i>Cynogale bennetti</i> EN	
Mammalia		<i>Hylobates funereus</i> EN	
Mammalia		<i>Lutra sumatrana</i> EN	
Mammalia		<i>Macaca fascicularis</i> EN	
Mammalia		<i>Macaca nemestrina</i> EN	
Mammalia		<i>Nasalis larvatus</i> EN	
Mammalia		<i>Pteropus</i> sp. EN	
Reptilia		<i>Cuora amboinensis</i> EN	
Aves		Vulnerable	<i>Buceros rhinoceros</i> VU
Aves			<i>Lophura ignita</i> VU
Mammalia	<i>Aonyx cinereus</i> VU		
Mammalia	<i>Maxomys whiteheadi</i> VU		
Mammalia	<i>Nycticebus menagensis</i> VU		
Reptilia	<i>Notochelys platynota</i> VU		
Aves	Near threatened		<i>Irena cyanogastra</i> NT

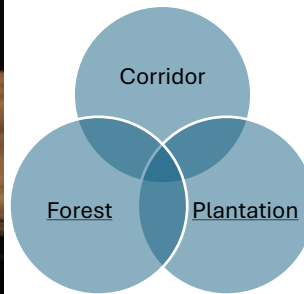
Figure 23 : Species detected and with a proven level of vulnerability according to the IUCN Red List



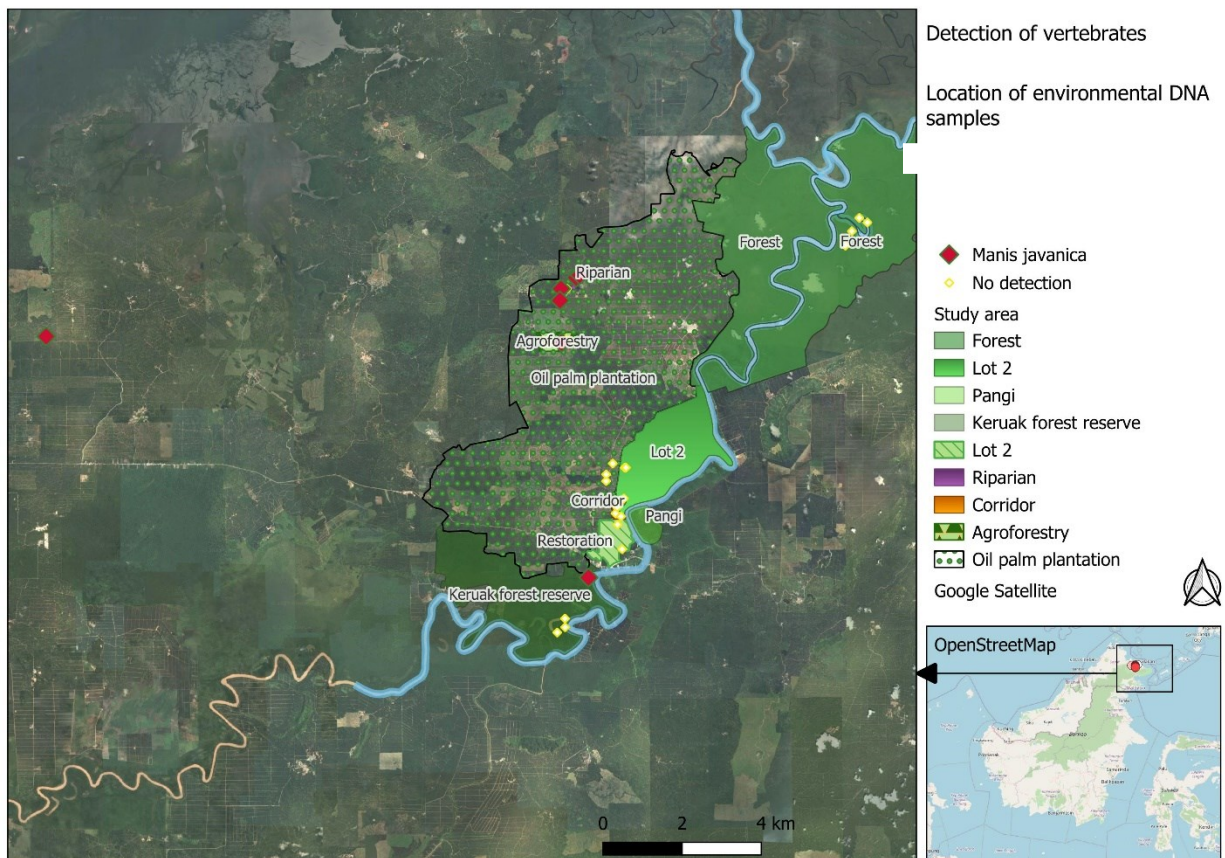
CLASS	Level Red List IUCN	Latin name
Mammalia	CR	<i>Manis javanica</i>



Sunda Pangolin



<https://www.iucnredlist.org/species/12763/123584856>

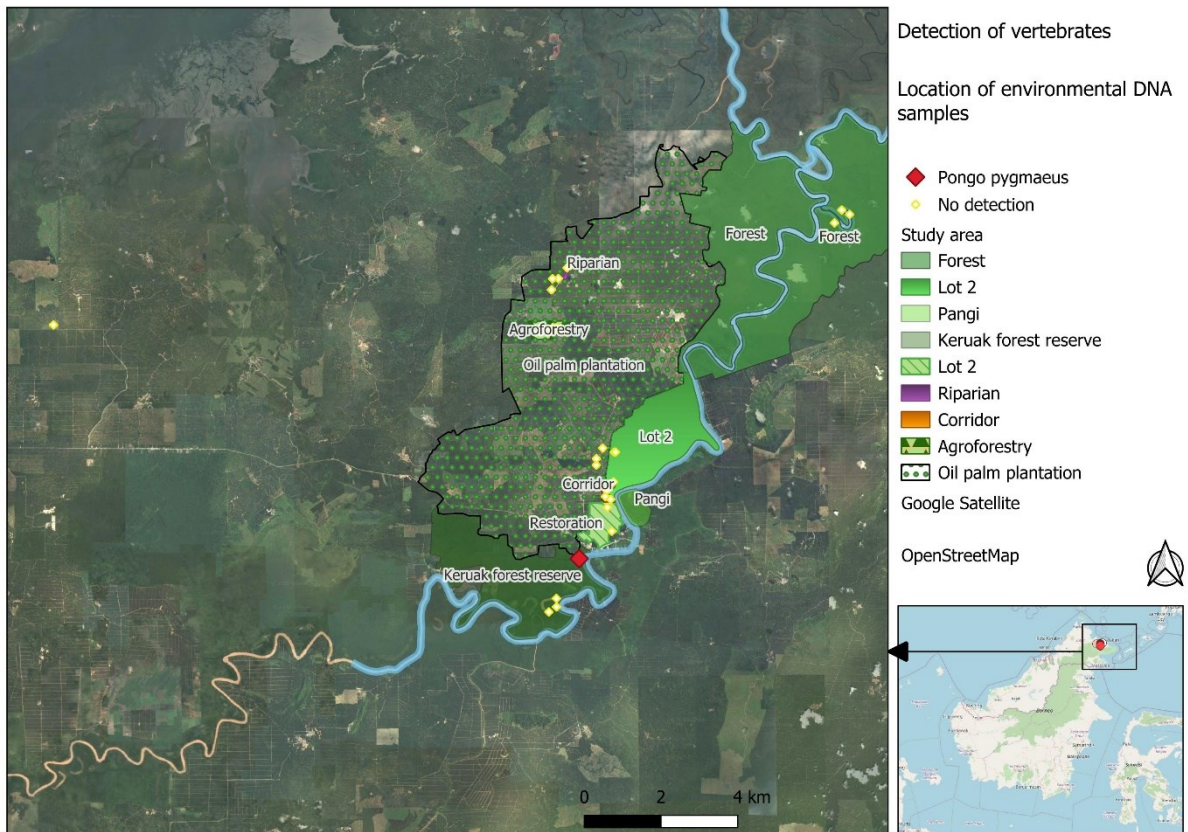
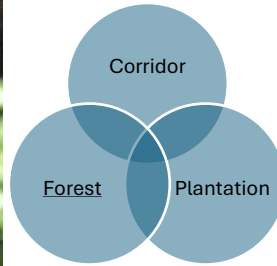


Present of 22% of the sites analysed, the species is found in forests and plantations.

CLASS	Level Red List IUCN	Latin name
Mammalia	CR	<i>Pongo pygmaeus</i>



Bornean Orangutan

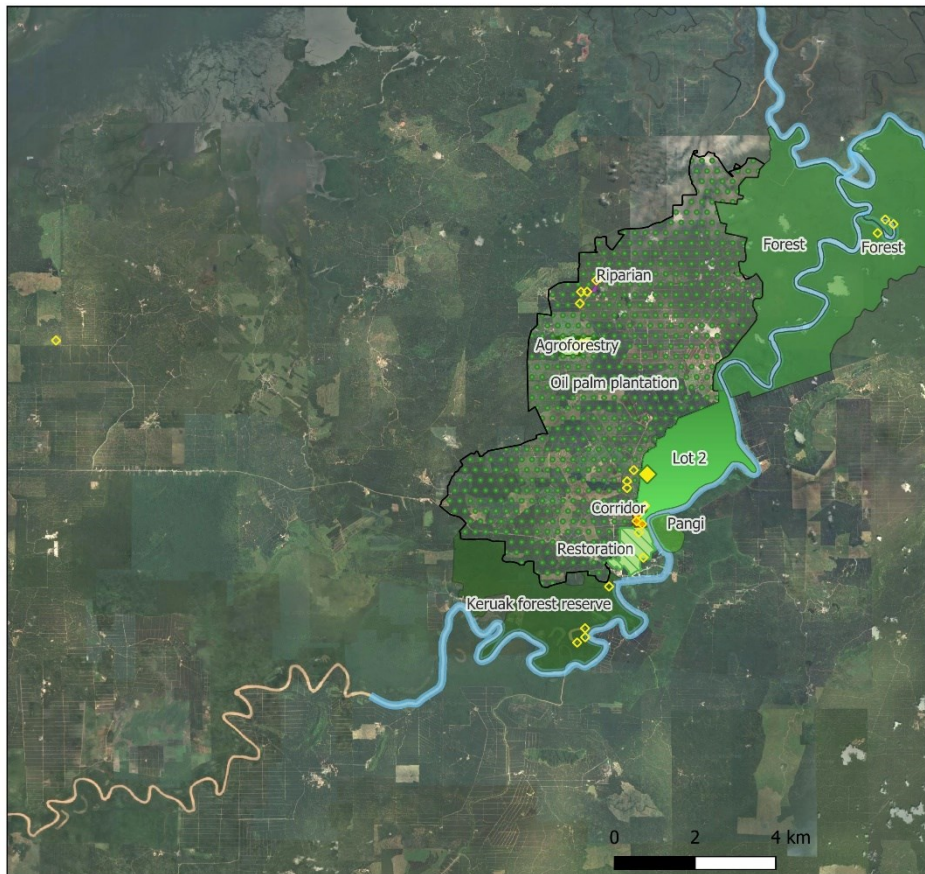
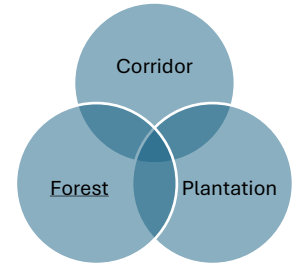


Species with at only 3% of the sites analysed.

CLASS	Level Red List IUCN	Latin name
Mammalia	EN	<i>Cynogale bennetti</i>



Otter Civet



Species with at only 3% of the sites analysed.

CLASS	Level Red List IUCN	Latin name
Mammalia	EN	<i>Hylobates funereus</i> EN



Northern Gray Gibbon

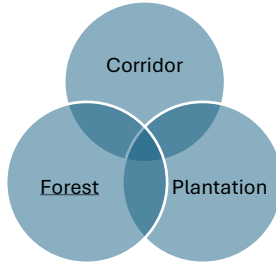
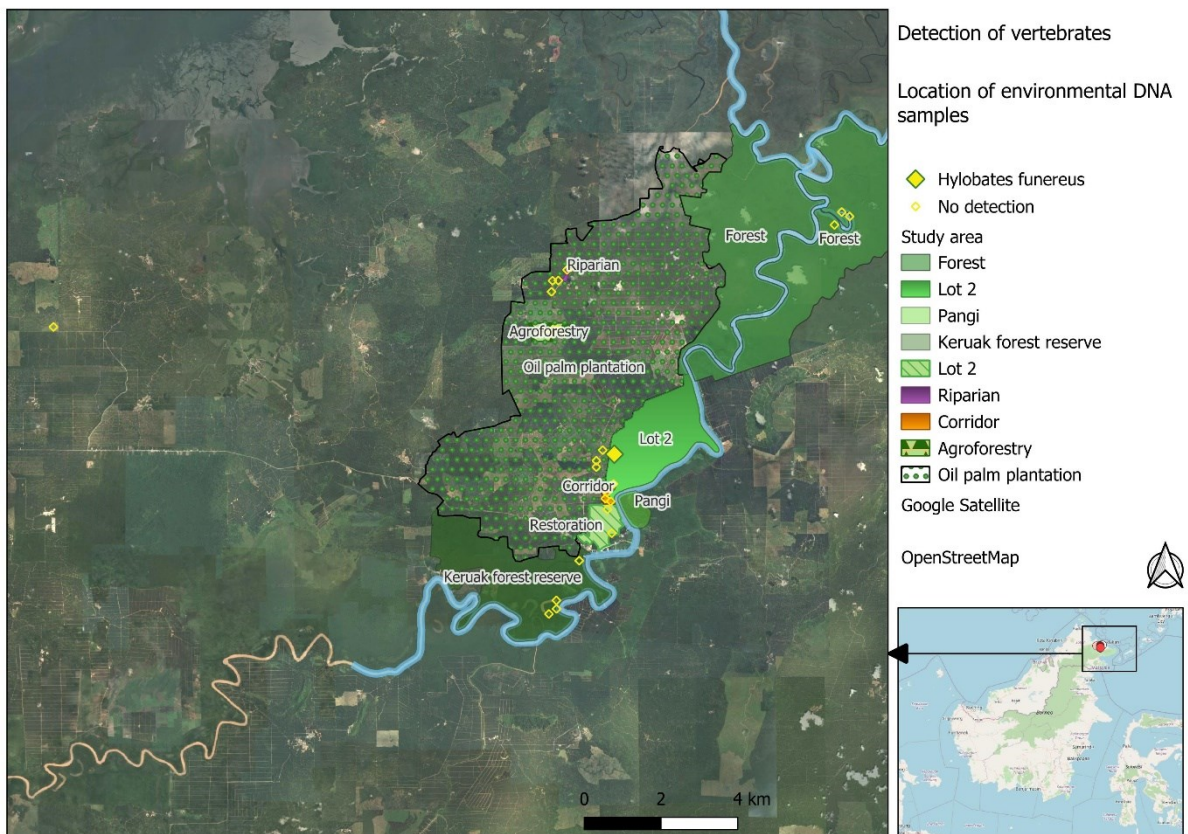


photo source wikipédia



Species with at only 3% of the sites analysed.

CLASS	Level Red List IUCN	Latin name
Mammalia	EN	<i>Lutra sumatrana</i> EN



Hairy-nosed Otter

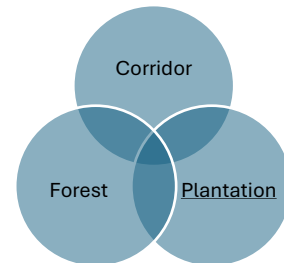
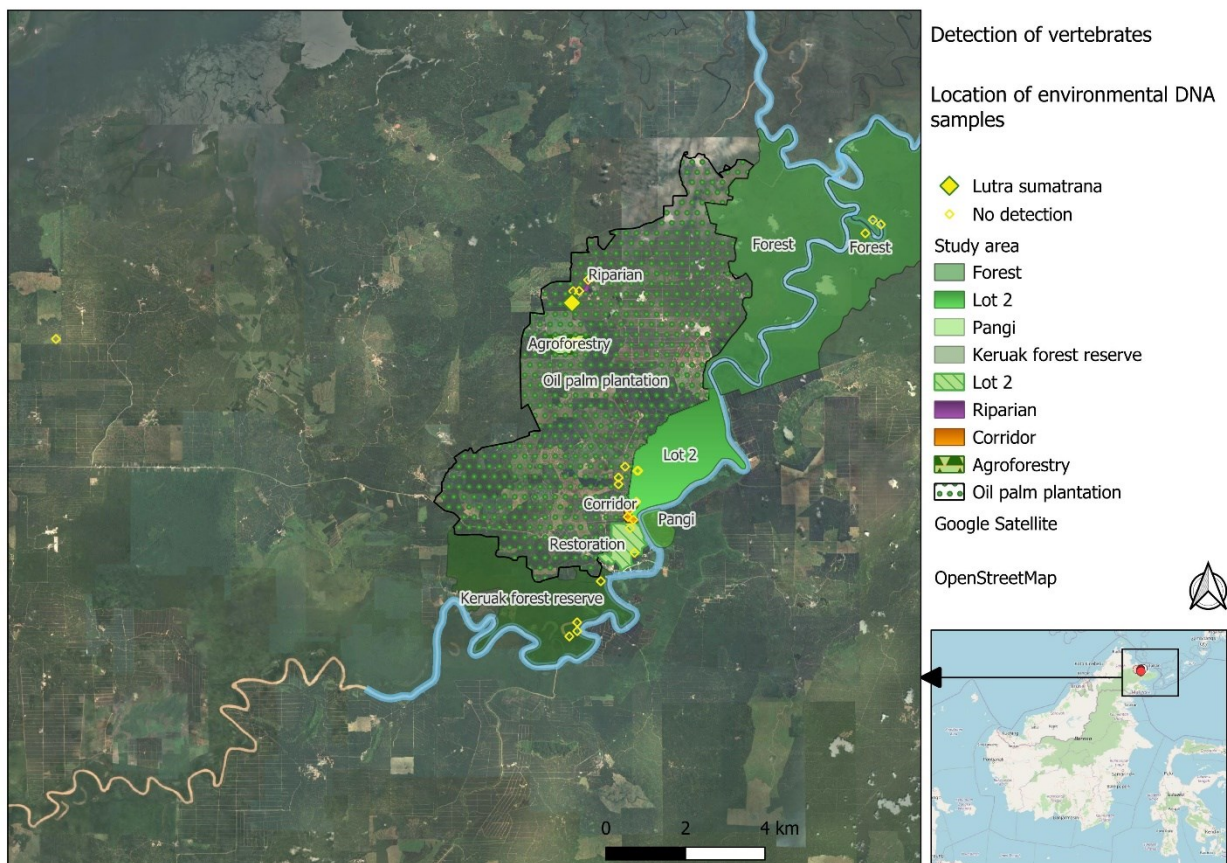


photo source wikipédia

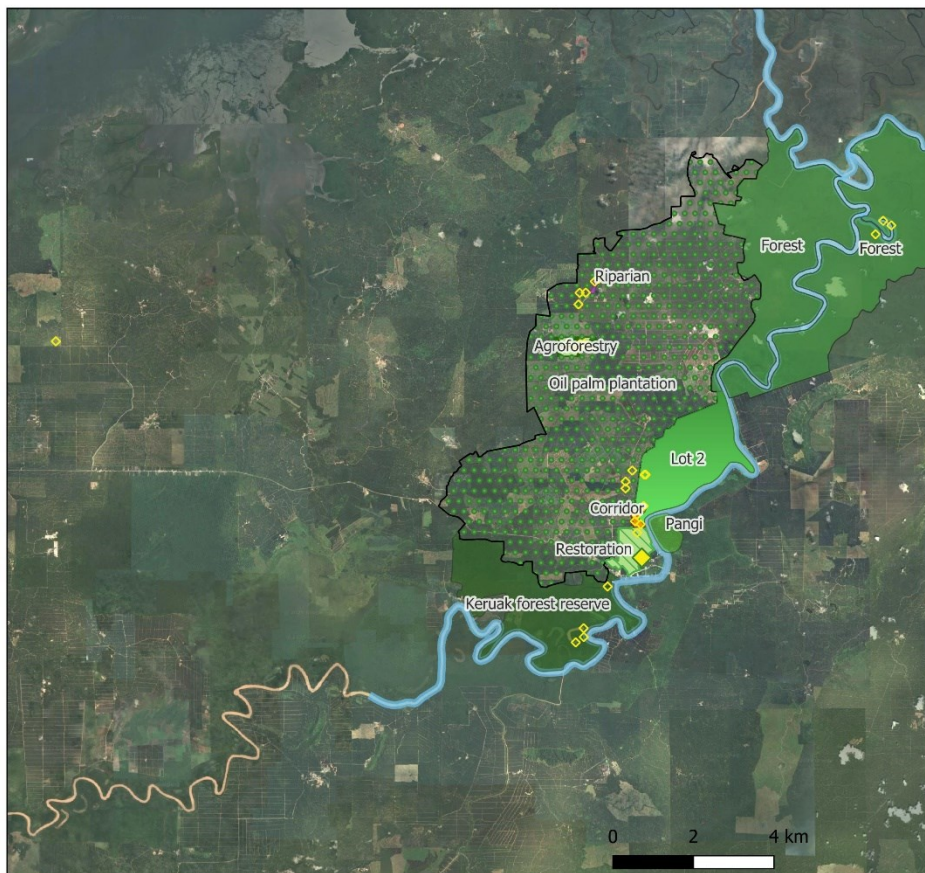
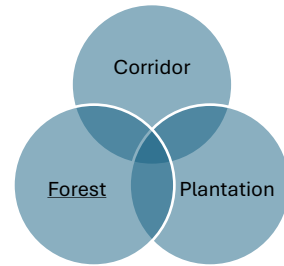


Species with at only 3% of the sites analysed.

CLASS	Level Red List IUCN	Latin name
Mammalia	EN	<i>Macaca fascicularis</i> EN



Common Long-tailed Macaque



Detection of vertebrates

Location of environmental DNA samples

- ◆ *Macaca fascicularis*
- ◇ No detection

Study area

- Forest
- Lot 2
- Pangli
- Keruak forest reserve
- Restoration
- Riparian
- Corridor
- Agroforestry
- Oil palm plantation

Google Satellite

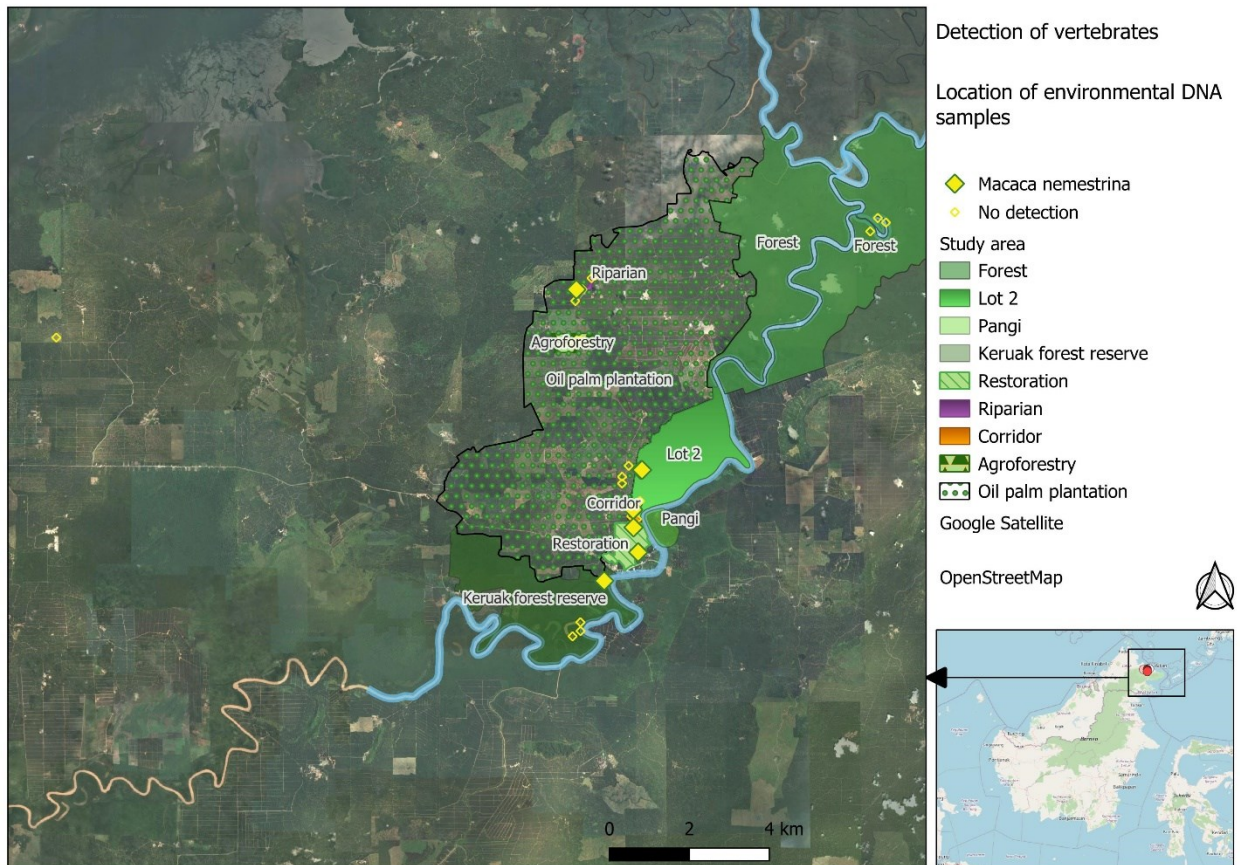
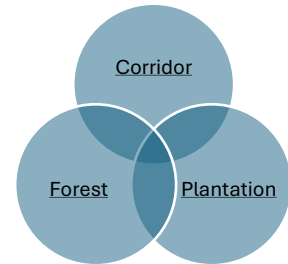
OpenStreetMap

Species with at only 3% of the sites analysed.

CLASSE	Level Red List IUCN	Latin name
Mammalia	EN	Macaca nemestrina EN



Southern Pig-tailed Macaque

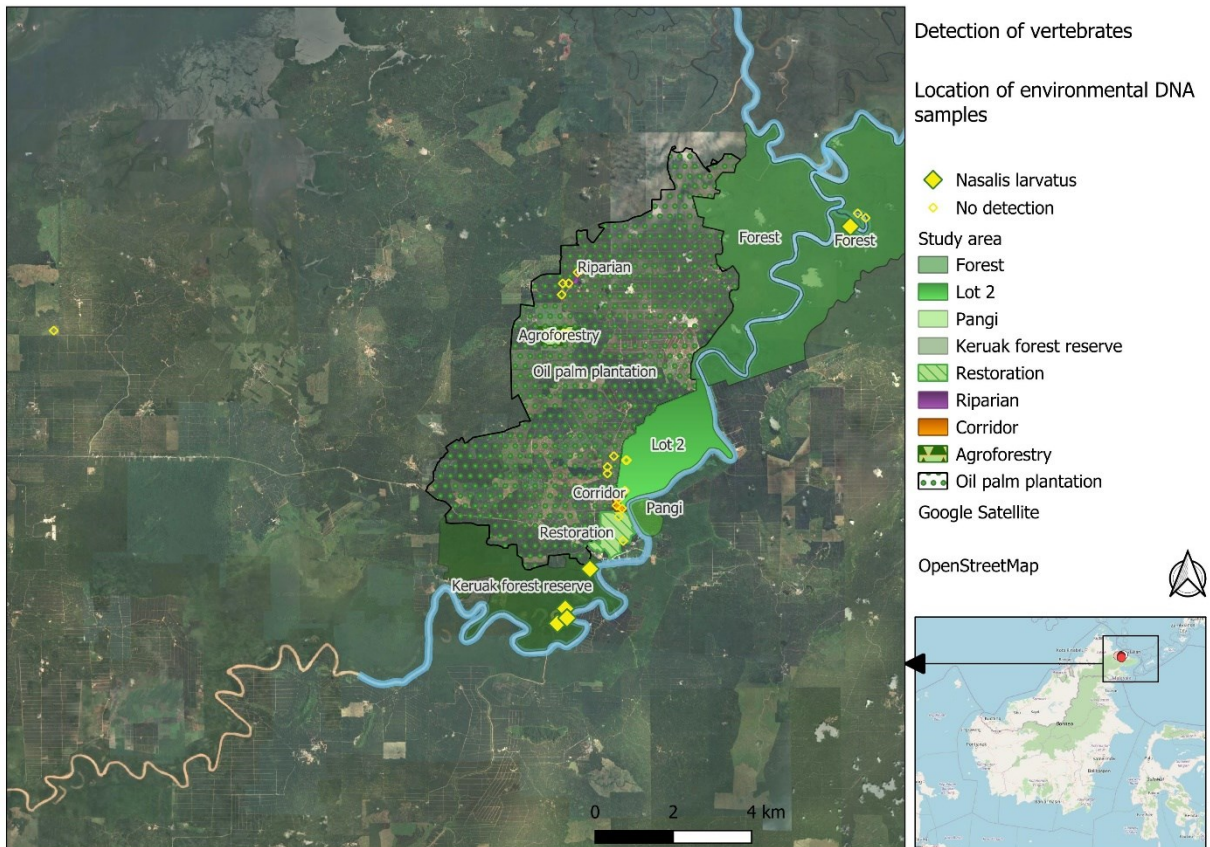
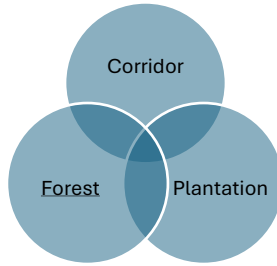


Species with at 22% of the sites analysed.

CLASS	Level Red List IUCN	Latin name
Mammalia	EN	<i>Nasalis larvatus</i> EN



Proboscis Monkey

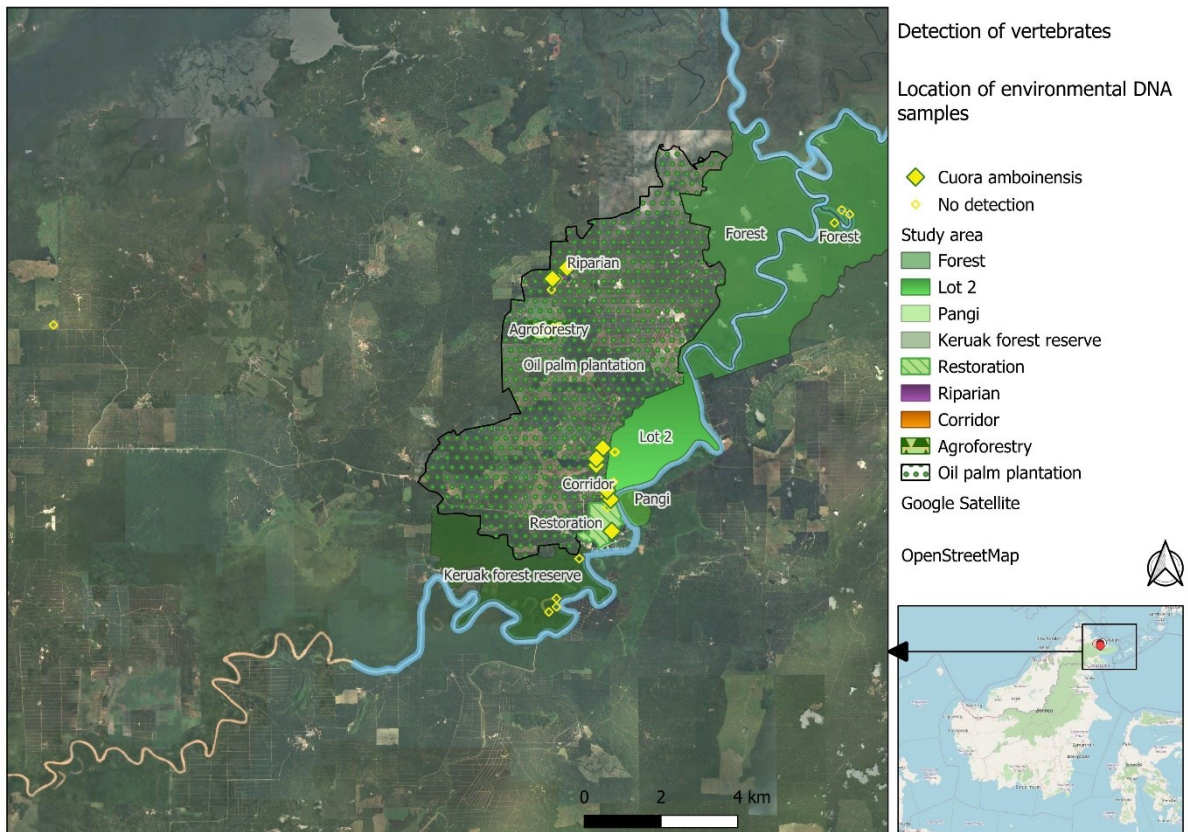
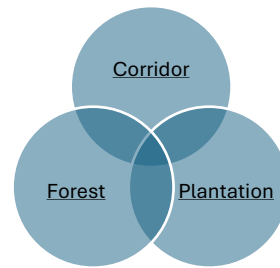


Species present at 22% of the sites analysed.

CLASS	Level Red List IUCN	Latin name
Reptilia	EN	<i>Cuora amboinensis</i> EN



Southeast Asian Box Turtle

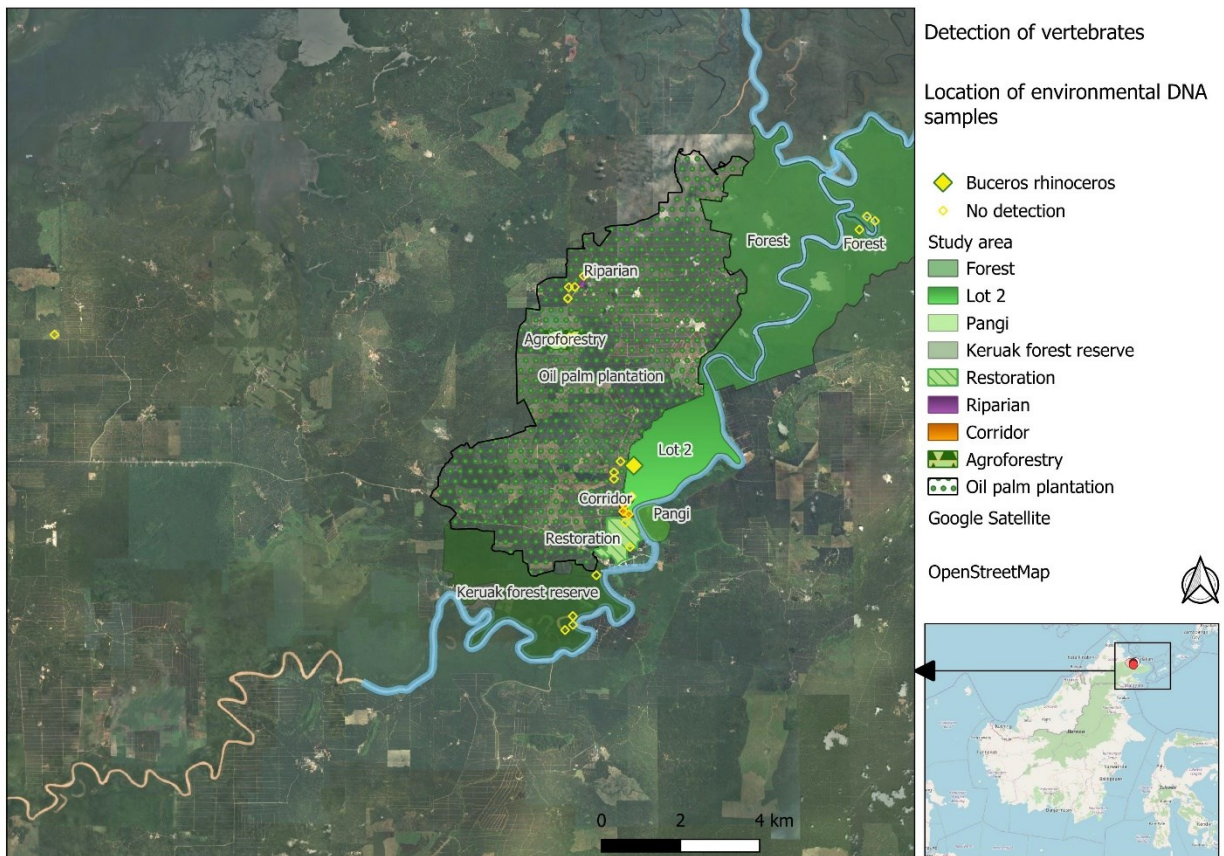
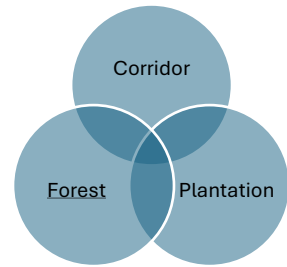


Species present at 25% of the sites analysed. This response for an aquatic reptile is quite strong, suggesting a relatively widespread presence of the species.

CLASS	Level Red List IUCN	Latin name
Aves	VU	<i>Buceros rhinoceros</i> VU



Rhinoceros Hornbill

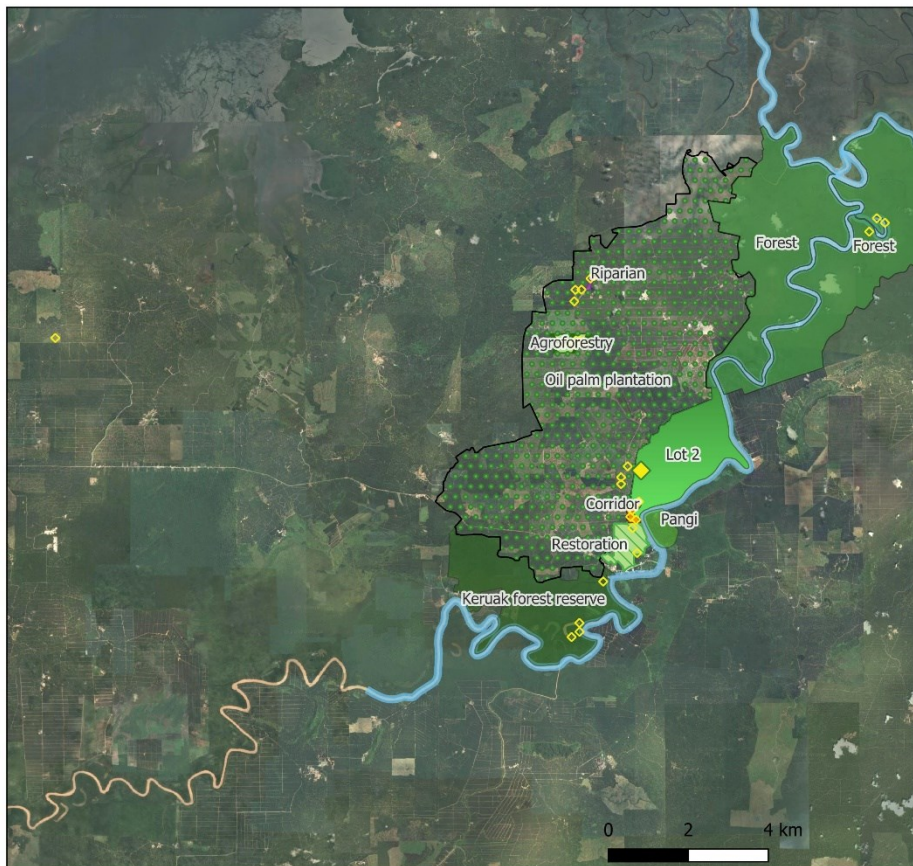
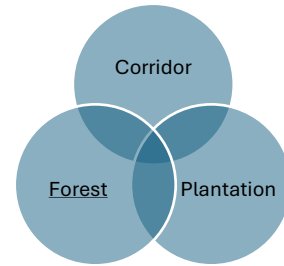


Species present at only 3% of the sites analysed. Bird detections are often underestimated for the bird class.

CLASS	Level Red List IUCN	Latin name
Aves	VU	<i>Lophura ignita</i> VU



Bornean Crested Fireback



Detection of vertebrates

Location of environmental DNA samples

- ◆ *Lophura ignita*
- ◇ No detection

Study area

- Forest
- Lot 2
- Pangli
- Keruak forest reserve
- Restoration
- Riparian
- Corridor
- Agroforestry
- Oil palm plantation

Google Satellite

OpenStreetMap

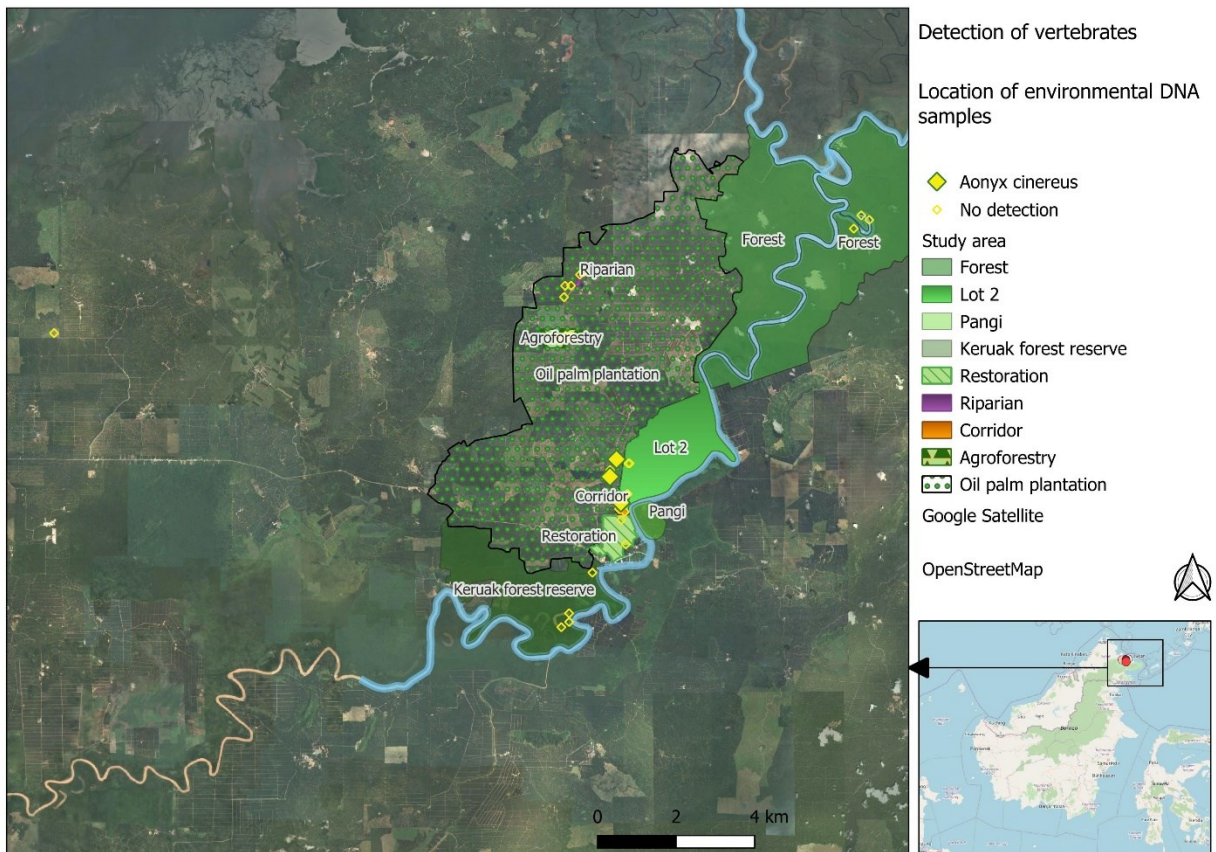
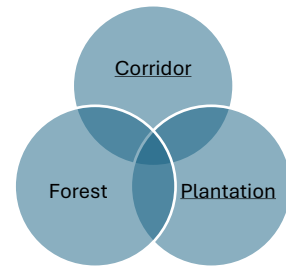


Species present in only 3% of the sites analysed. Bird detections are often underestimated for the bird class.

CLASS	Level Red List IUCN	Latin name
Mammalia	VU	<i>Aonyx cinereus</i> VU



Asian Small-clawed Otter

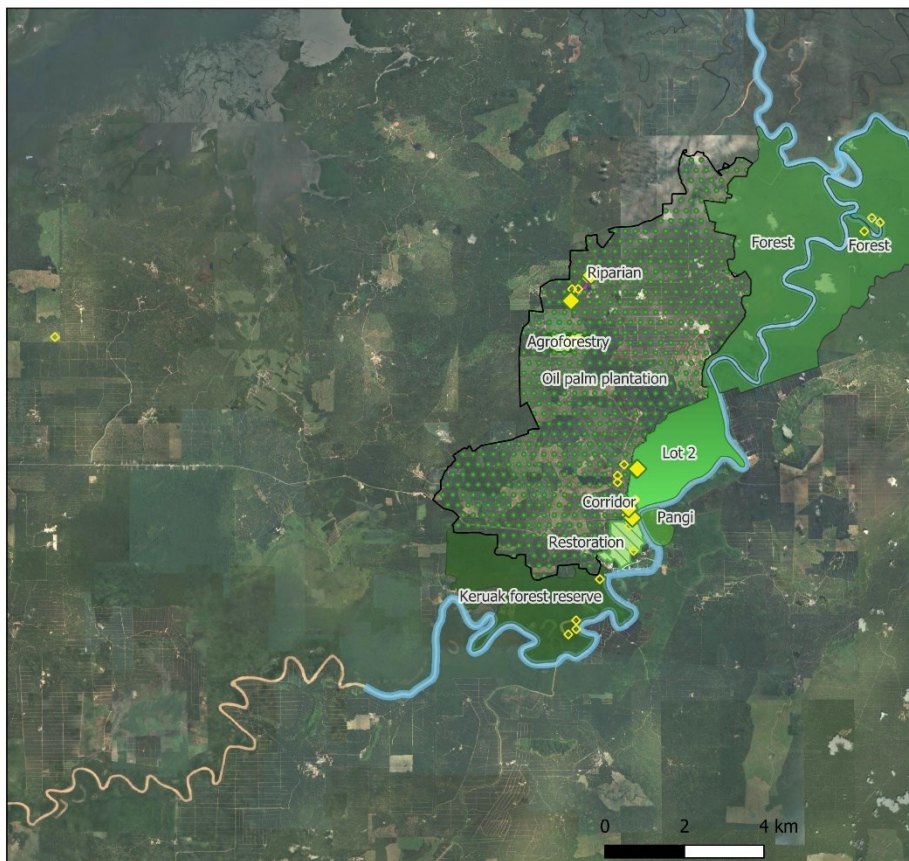
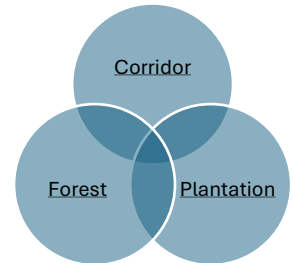


Species present at only 9% of the sites analysed.

CLASS	Level Red List IUCN	Latin name
Mammalia	VU	<i>Maxomys whiteheadi</i> VU



Whitehead's Sundaic Maxomys

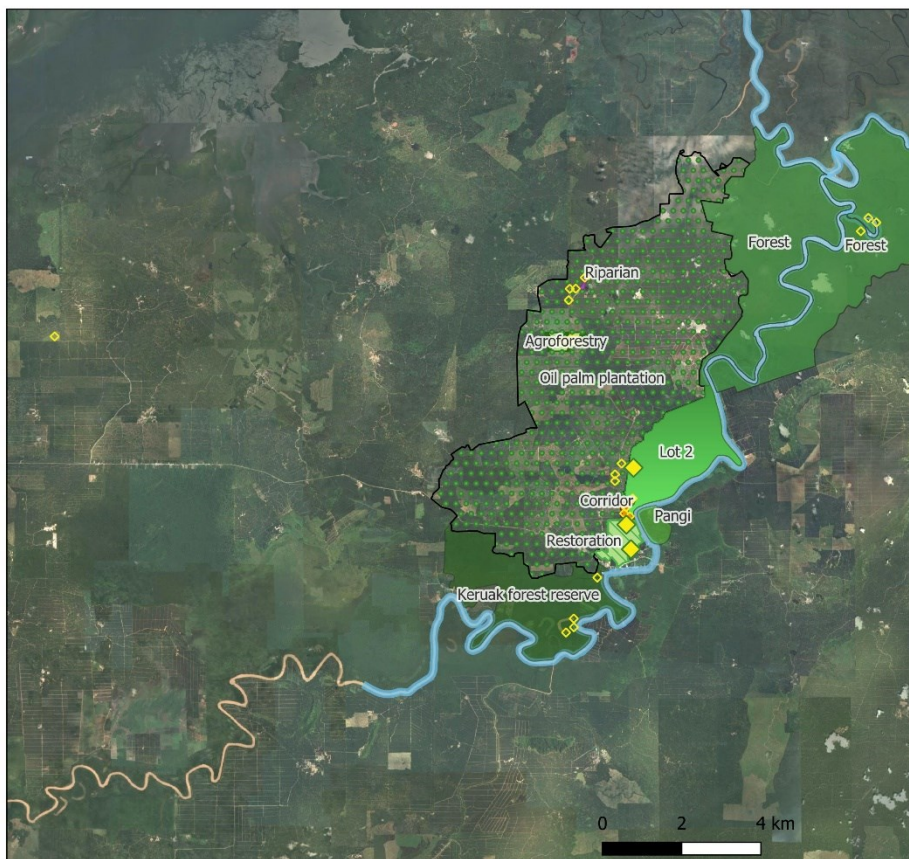
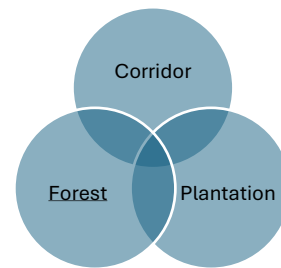


Species present at 19% of the sites analysed.

CLASS	Level Red List IUCN	Latin name
Mammalia	VU	<i>Nycticebus menagensis</i> VU



Philippine Slow Loris

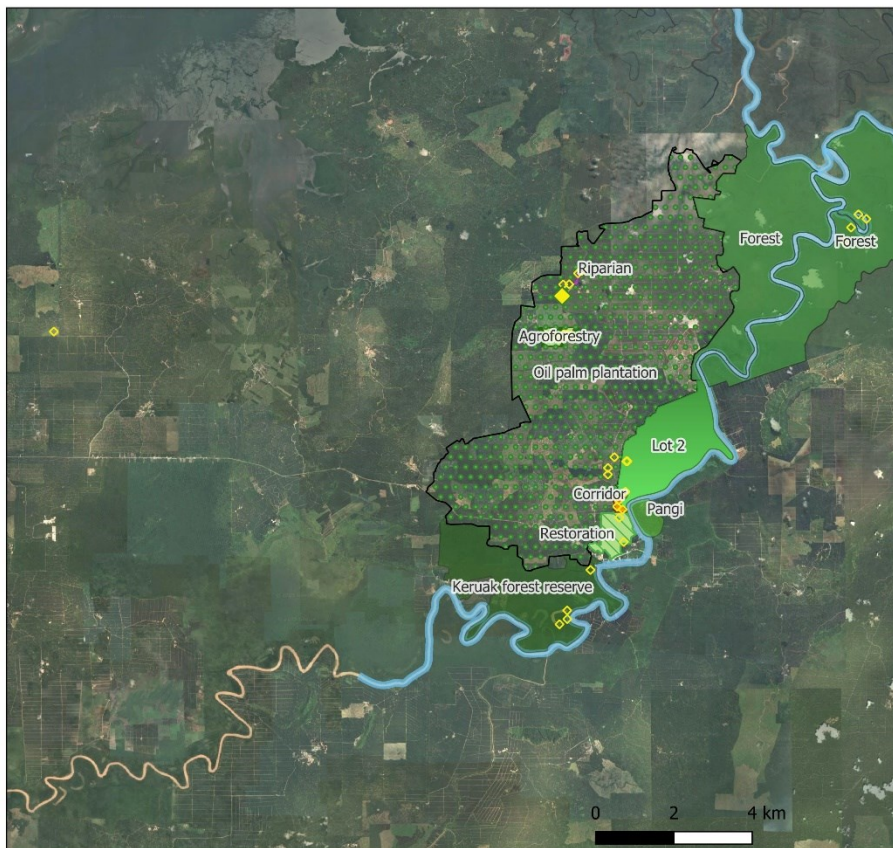
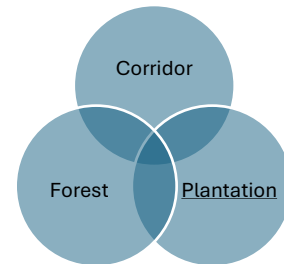


Species present at 9% of the sites analysed.

CLASS	Level Red List IUCN	Latin name
Reptilia	VU	<i>Notochelys platynota</i> VU



Malayan Flat-shelled Turtle



Detection of vertebrates

Location of environmental DNA samples

- ◆ *Notochelys platynota*
- ◇ No detection

- Study area
- Forest
- Lot 2
- Pangi
- Keruak forest reserve
- Restoration
- Riparian
- Corridor
- Agroforestry
- Oil palm plantation

Google Satellite

OpenStreetMap

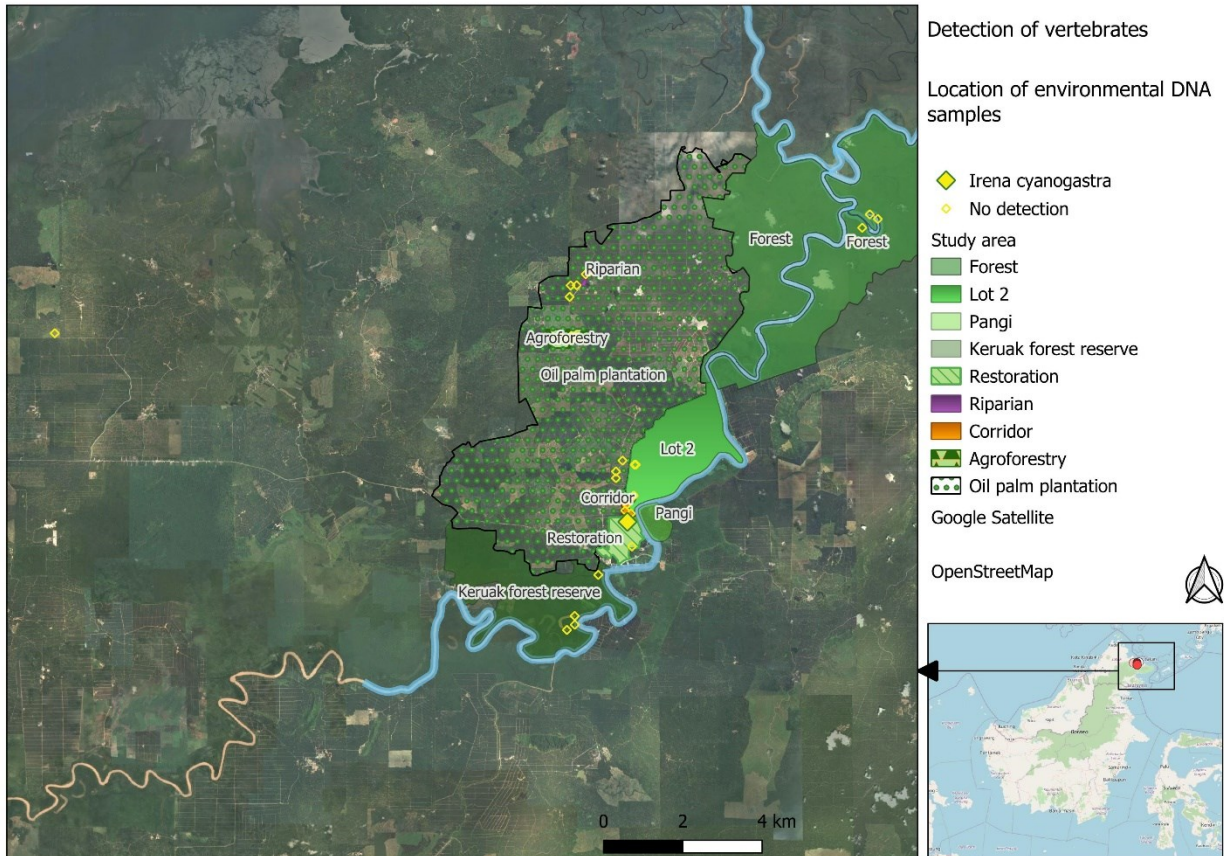
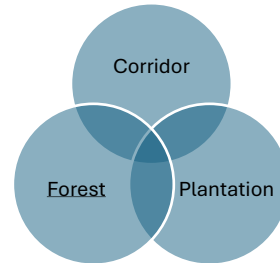


Species present at only 3% of the sites analysed.

CLASS	Level Red List IUCN	Latin name
Aves	NT	<i>Irena cyanogastra</i> NT



Philippine Fairy-bluebird



Species present at 3% of the sites analysed. As a bird species, it is highly likely that detection by eDNA is underestimated for this species.

Comparison between two palm groves of different ages

Here we compare the diversity of an old plantation with a young plantation.

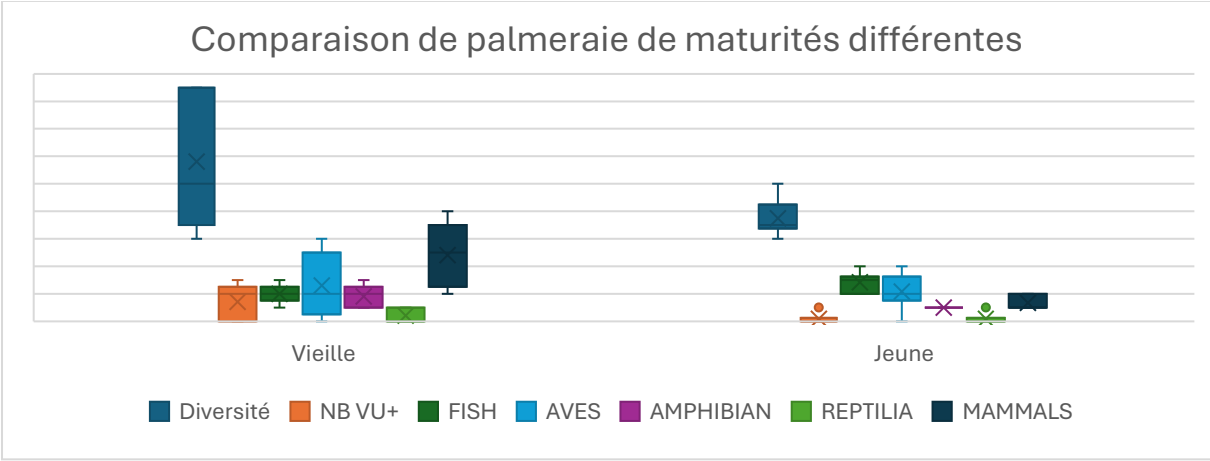


Figure 24 : Comparison between two palm groves of different ages

As expected, the **specific richness** is greater for **ancient palm groves**.

This translates into a significant increase in the number of mammal and amphibian species. To a lesser extent, bird diversity tends to increase with the age of the palm grove.

In terms of conservation, older palm groves are home to more vulnerable species than younger ones. This finding has important implications for managing new palm rotation cycles. Rotation cycles span approximately 25 years, and in order to fulfil the various certification requirements, land-use planning for a new rotation may strategically designate certain areas as High Conservation Value Areas that will no longer be used for palm production. In these areas, the restoration approach should involve retaining the existing mature palm plants rather than felling them, while establishing new tree seedlings in the understory beneath the palm canopy.

Factor	older plantation	Younger plantation
Overall diversity	12+-7	8+-1
Vulnerable species	1.4	0.166
Fish diversity	2	2.8
Bird diversity	2.6	2
Amphibian diversity	1.8	1
Reptile diversity	0.4	0.1
Mammal diversity	4.8	1.3
	++	--

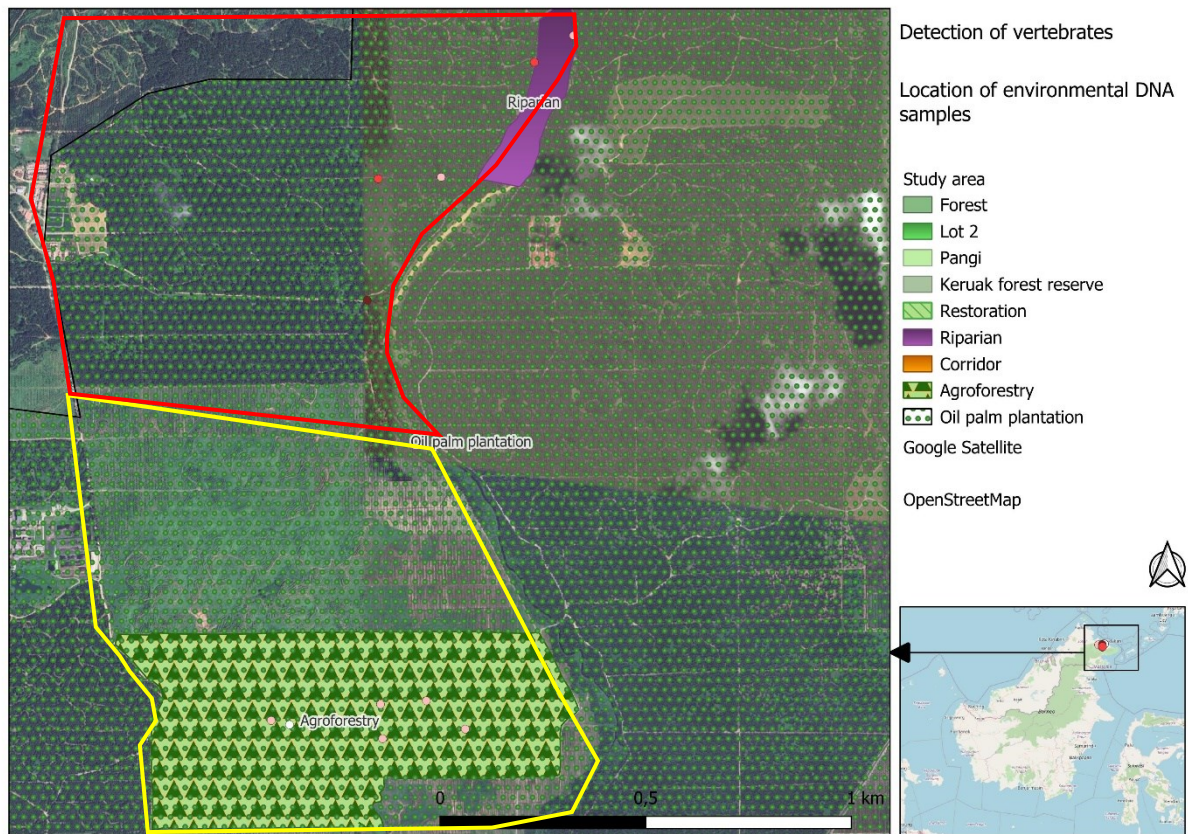


Figure 25 : Map showing the location of samples from two palm groves of different ages

In figure 25, younger palm groves are located in the south and the older ones in the north.

The "Young Palms" sites are part of an agroforestry experiment called TRAILS, where native trees and oil palms are intercropped and planted together. This experiment began in 2022, meaning both the trees and palms are still very young and do not yet support the same level of biodiversity as mature palm stands. However, the data collected during this eDNA campaign provides critical baseline information captured at a pivotal moment—shortly after this habitat underwent a massive transformation through the removal of old mature palms and the establishment of a new rotation cycle integrating both palms and trees. Long-term monitoring by Hutan will reveal how biodiversity in these agroforestry systems compares to that in conventional monoculture oil palm plantations.

Comparison between two palm groves of different ages and an ecological corridor

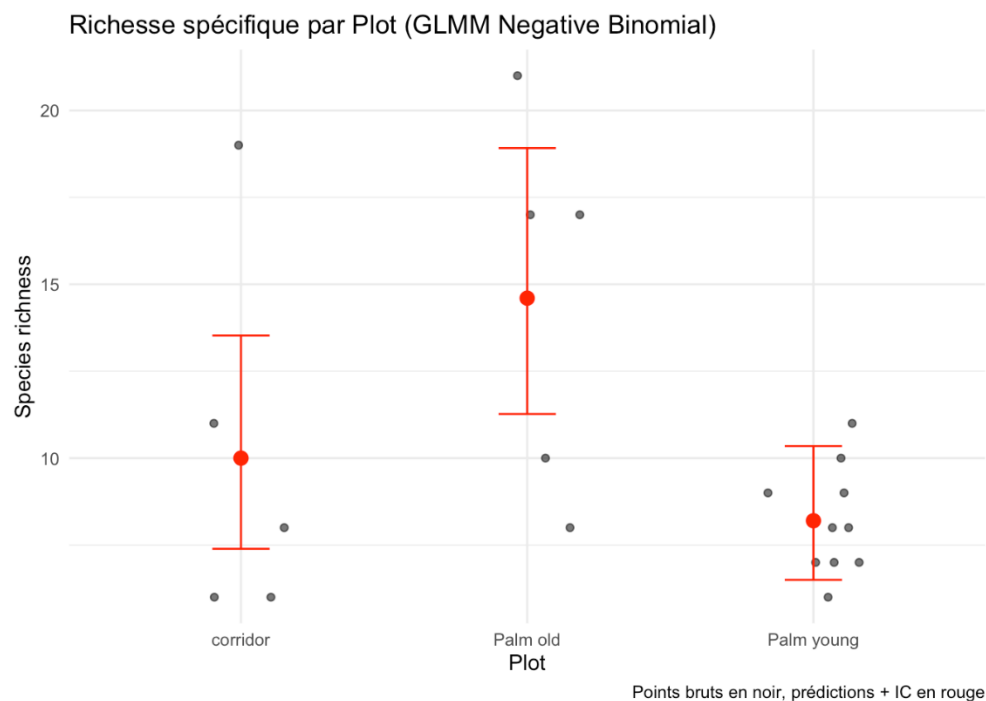


Figure 26 : Specific richness per plot

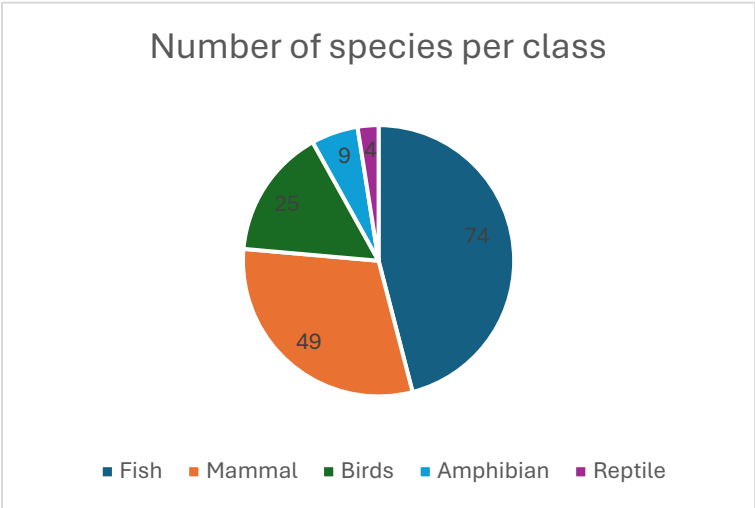
There is a significant difference between the richness specific of old and young palms. The diversity is most important in old palms (see above). For the corridor, the difference between old and young palm is less significant.

The results reveal high variability in both the Corridor and mature palm sites, in contrast to the remarkably low variability observed in the young agroforestry experiment. This finding is significant for several reasons:

- **Sampling bias in biodiversity assessments:** Due to the high spatial variability within the corridor and mature palm habitats, biodiversity monitoring results will be strongly influenced by survey plot placement. This factor is rarely accounted for in oil palm plantation biodiversity research, potentially leading to inconsistent or misleading conclusions.
- **Need for mechanistic understanding:** Further investigation is needed to identify the drivers of this high variability, such as proximity to forest edges, understory vegetation quality, management practices, human interventions and other factors...
- **Baseline conditions following rotation:** The exceptionally low variability in the young palms within the TRAILS experiment (n=10 samples) reflects the intensive habitat transformation that occurred at the end of the first rotation cycle (approximately one year before replanting). These results demonstrate that the transition to a new rotation effectively eliminates nearly all biodiversity, resetting the habitat to an extremely degraded baseline from which ecological recovery must begin.

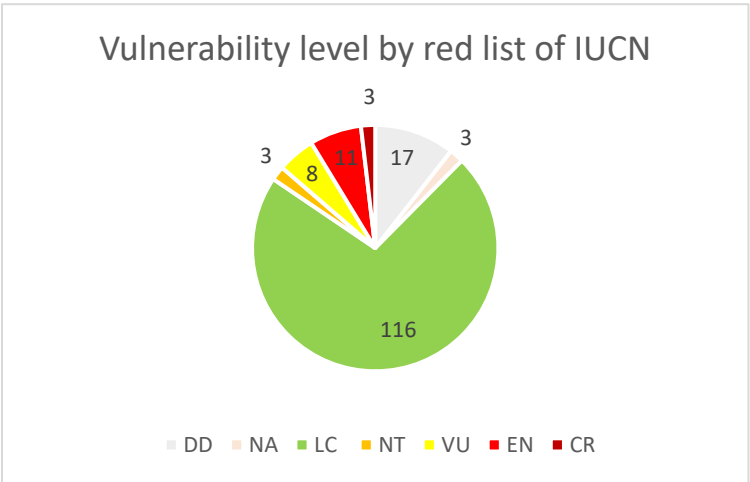
Summary of eDNA data since 2019 on the Kinabatangan

During our first campaign in 2019 we detected **160** species, mostly fishes since we collected water in the rivers, ditch, drain and oxbow lakes.



Mammals was the second most common class with 49 species followed by birds (24 sp.), amphibians (9 sp.) and reptiles (4 sp.).

Figure 27 : Number of species per class since 2019



In this list, 16% species are endangered but we need to acknowledge that the status of several species (especially fishes) has not been assessed yet assessment, and some are unknown in genetic databases. the red list is not completely up to date.

Figure 28 : Vulnerability level by red list of IUCN

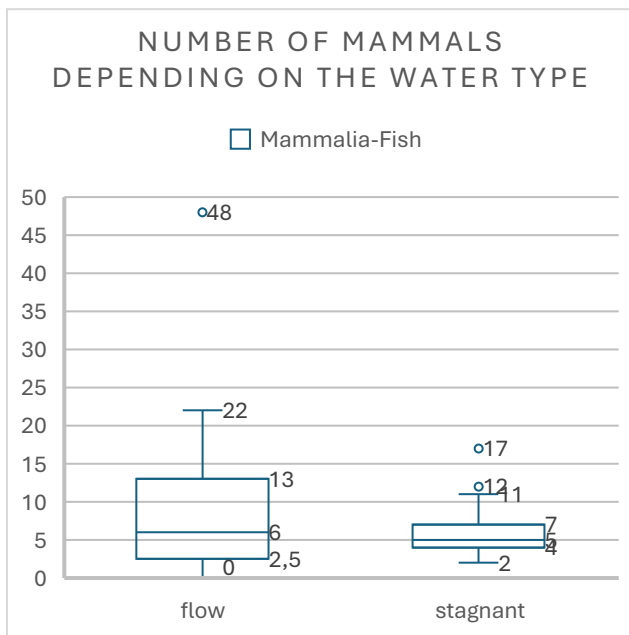


Figure 29 : Number of mammals depending on the water type

We now compare the fish community; we strongly advise to use the flowing water eDNA approach.

eDNA results in stagnant water (puddles, ponds, oxbow lakes: second campaign) and flowing water (drain, ditch, river).

Overall, results seem more complete when we follow the eDNA flowing water protocol. This result is not surprising since more freshwater biodiversity is expected to be found in large flowing water. However more terrestrial species were recorded with the stagnant water samples. So the choice of running vs stagnant water sampling will primarily depend on the questions we want

to investigate. For the fish community, we strongly advise using the flowing water eDNA sampling approach.

However, undertaking this methodology is more troublesome than using the stagnant water approach.

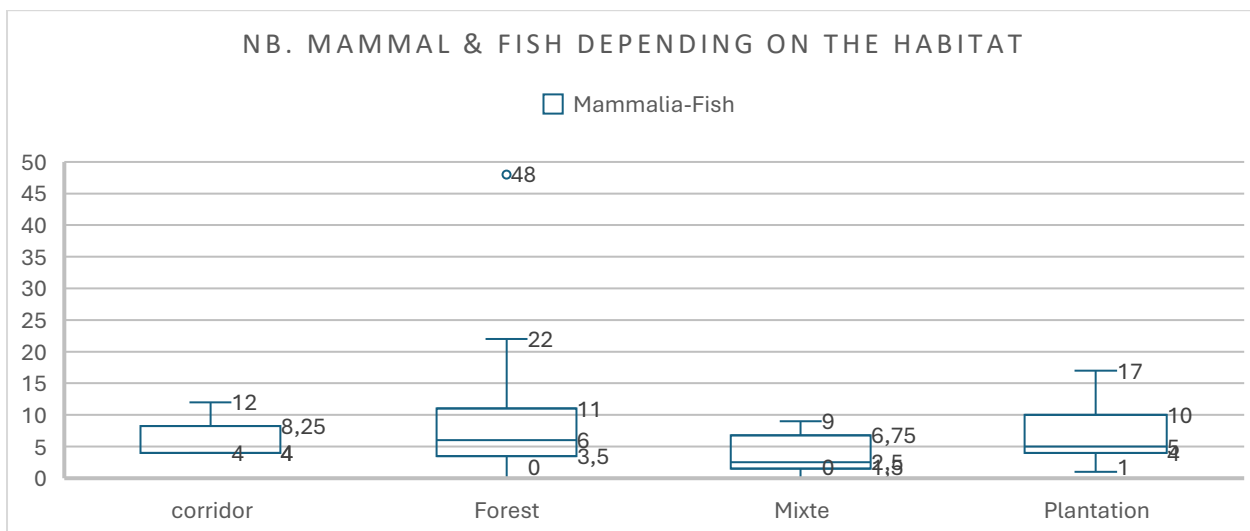


Figure 30 : Number mammal and fish depending on the habitat

As expected, more species were overall identified in the forest habitat.

Conclusion

This second collection campaign aimed to:

- Compare an approach using stagnant water and running water,
- Contribute to the biodiversity monitoring carried out by Hutan,
- Participate in identifying species present in different environments,
- Provide an opportunity to train Hutan teams in new technologies.

The first campaign in stagnant water identified several new species, including mammals, birds, fish and reptiles.

In total, the eDNA campaigns identified 161 species, including 20 new detections for HUTAN in 2019.

In conclusion, the eDNA approach made it possible to identify certain species that had not yet been detected by Hutan teams during successive inventories, thus demonstrating the value of this approach for detecting rare or cryptic species.

These environmental DNA results will be integrated into Hutan's monitoring data to complete the picture of biodiversity in the different environments of the Kinabatangan.



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Annexes

Species	Class	Order	Family	Conservation
<i>Acentrogobius janthinopterus</i>	Actinopteri	Gobiiformes	Gobiidae	LC
<i>Acentrogobius viridipunctatus</i>	Actinopteri	Gobiiformes	Gobiidae	LC
<i>Alectis indica</i>	Actinopteri	Carangiformes	Carangidae	LC
<i>Alepes djedaba</i>	Actinopteri	Carangiformes	Carangidae	LC
<i>Anabas testudineus</i>	Actinopteri	Anabantiformes	Anabantidae	LC
<i>Anguilla malgumora</i>	Actinopteri	Anguilliformes	Anguillidae	DD
<i>Anodontostoma chacunda</i>	Actinopteri	Clupeiformes	Clupeidae	LC
<i>Apogon hyalosoma</i>	Actinopteri	Kurtiformes	Apogonidae	LC
<i>Barbonymus gonionotus</i>	Actinopteri	Cypriniformes	Cyprinidae	LC
<i>Boleophthalmus boddarti</i>	Actinopteri	Gobiiformes	Gobiidae	LC
<i>Brachygobius doriae</i>	Actinopteri	Gobiiformes	Gobiidae	LC
<i>Butis koilomatodon</i>	Actinopteri	Gobiiformes	Eleotridae	LC
<i>Caranx ignobilis</i>	Actinopteri	Carangiformes	Carangidae	LC
<i>Channa striata</i>	Actinopteri	Anabantiformes	Channidae	LC
<i>Chanos chanos</i>	Actinopteri	Gonorynchiformes	Chanidae	LC
<i>Chitala ornata</i>	Actinopteri	Osteoglossiformes	Notopteridae	LC
<i>Decapterus macrosoma</i>	Actinopteri	Carangiformes	Carangidae	LC
<i>Decapterus maruadsi</i>	Actinopteri	Carangiformes	Carangidae	LC
<i>Drombus kranjiensis</i>	Actinopteri	Gobiiformes	Gobiidae	DD
<i>Eleotris fusca</i>	Actinopteri	Gobiiformes	Eleotridae	LC
<i>Ellochelon vaigiensis</i>	Actinopteri	Mugiliformes	Mugilidae	LC
<i>Escualosa thoracata</i>	Actinopteri	Clupeiformes	Clupeidae	LC
<i>Eugnathogobius variegatus</i>	Actinopteri	Gobiiformes	Gobiidae	LC
<i>Gerres filamentosus</i>	Actinopteri	Gerreiformes	Gerreidae	LC
<i>Glossogobius aureus</i>	Actinopteri	Gobiiformes	Gobiidae	LC
<i>Glossogobius giuris</i>	Actinopteri	Gobiiformes	Gobiidae	LC

<i>Hemigobius hoevenii</i>	Actinopteri	Gobiiformes	Gobiidae	LC
<i>Hemigobius mingi</i>	Actinopteri	Gobiiformes	Gobiidae	DD
<i>Hemiramphus far</i>	Actinopteri	Beloniformes	Hemiramphidae	no
<i>Hyporhamphus limbatus</i>	Actinopteri	Beloniformes	Hemiramphidae	LC
<i>Hypostomus plecostomus</i>	Actinopteri	Siluriformes	Loricariidae	no
<i>Jaydia novaeguineae</i>	Actinopteri	Kurtiformes	Apogonidae	no
<i>Johnius belangerii</i>	Actinopteri	-	Sciaenidae	LC
<i>Lagocephalus lunaris</i>	Actinopteri	Tetraodontiformes	Tetraodontidae	LC
<i>Lethrinus lentjan</i>	Actinopteri	Spariformes	Lethrinidae	LC
<i>Luciosoma bleekeri</i>	Actinopteri	Cypriniformes	Danionidae	LC
<i>Lutjanus argentimaculatus</i>	Actinopteri	Lutjaniformes	Lutjanidae	LC
<i>Lutjanus johnii</i>	Actinopteri	Lutjaniformes	Lutjanidae	LC
<i>Lutjanus notatus</i>	Actinopteri	Lutjaniformes	Lutjanidae	LC
<i>Lutjanus russellii</i>	Actinopteri	Lutjaniformes	Lutjanidae	LC
<i>Megalops cyprinoides</i>	Actinopteri	Elopiformes	Megalopidae	DD
<i>Monodactylus argenteus</i>	Actinopteri	-	Monodactylidae	LC
<i>Moolgarda engeli</i>	Actinopteri	Mugiliformes	Mugilidae	LC
<i>Moolgarda perusii</i>	Actinopteri	Mugiliformes	Mugilidae	LC
<i>Mugilogobius chulae</i>	Actinopteri	Gobiiformes	Gobiidae	LC
<i>Ophiocara porocephala</i>	Actinopteri	Gobiiformes	Eleotridae	LC
<i>Oxyeleotris marmorata</i>	Actinopteri	Gobiiformes	Eleotridae	LC
<i>Pangasianodon hypophthalmus</i>	Actinopteri	Siluriformes	Pangasiidae	EN
<i>Parapocryptes serperaster</i>	Actinopteri	Gobiiformes	Gobiidae	LC
<i>Pellona ditchela</i>	Actinopteri	Clupeiformes	Pristigasteridae	LC
<i>Periophthalmodon schlosseri</i>	Actinopteri	Gobiiformes	Gobiidae	LC
<i>Periophthalmus argentilineatus</i>	Actinopteri	Gobiiformes	Gobiidae	LC
<i>Periophthalmus novemradiatus</i>	Actinopteri	Gobiiformes	Gobiidae	DD
<i>Photopectoralis bindus</i>	Actinopteri	Chaetodontiformes	Leiognathidae	DD
<i>Pisodonophis boro</i>	Actinopteri	Anguilliformes	Ophichthidae	LC
<i>Planiliza macrolepis</i>	Actinopteri	Mugiliformes	Mugilidae	LC
<i>Planiliza subviridis</i>	Actinopteri	Mugiliformes	Mugilidae	LC
<i>Plicofollis argyropleuron</i>	Actinopteri	Siluriformes	Ariidae	no
<i>Pomadasys hasta</i>	Actinopteri	Lutjaniformes	Haemulidae	LC
<i>Psammogobius biocellatus</i>	Actinopteri	Gobiiformes	Gobiidae	LC
<i>Pseudogobius javanicus</i>	Actinopteri	Gobiiformes	Gobiidae	no
<i>Puntioplites proctozystron</i>	Actinopteri	Cypriniformes	Cyprinidae	LC
<i>Rasbora hobelmani</i>	Actinopteri	Cypriniformes	Danionidae	NA
<i>Rasbora steineri</i>	Actinopteri	Cypriniformes	Cyprinidae	LC
<i>Rastrelliger kanagurta</i>	Actinopteri	Scombriformes	Scombridae	DD
<i>Scatophagus argus</i>	Actinopteri	-	Scatophagidae	LC
<i>Scomber scombrus</i>	Actinopteri	Scombriformes	Scombridae	LC
<i>Selar crumenophthalmus</i>	Actinopteri	Carangiformes	Carangidae	LC
<i>Setipinna melanochir</i>	Actinopteri	Clupeiformes	Engraulidae	DD
<i>Siganus guttatus</i>	Actinopteri	-	Siganidae	LC
<i>Sillago sihama</i>	Actinopteri	-	Sillaginidae	LC
<i>Tetraodon nigroviridis</i>	Actinopteri	Tetraodontiformes	Tetraodontidae	no

<i>Toxotes chatareus</i>	Actinopteri	(vide)	Toxotidae	LC
<i>Trichopodus pectoralis</i>	Actinopteri	Anabantiformes	Osphronemidae	LC
<i>Chaperina fusca</i>	Amphibia	Anura	Microhylidae	LC
<i>Hylarana chalconota</i>	Amphibia	Anura	Ranidae	LC
<i>Hylarana erythraea</i>	Amphibia	Anura	Ranidae	LC
<i>Ingerophrynus divergens</i>	Amphibia	Anura	Bufoidea	LC
<i>Kurixalus appendiculatus</i>	Amphibia	Anura	Rhacophoridae	LC
<i>Limnonectes finchi</i>	Amphibia	Anura	Dicroglossidae	LC
<i>Limnonectes ingeri</i>	Amphibia	Anura	Dicroglossidae	LC
<i>Limnonectes kuhlii species complex</i>	Amphibia	Anura	Dicroglossidae	LC
<i>Polypedates leucomystax</i>	Amphibia	Anura	Rhacophoridae	LC
<i>Acridotheres cristatellus</i>	Aves	Passeriformes	Sturnidae	LC
<i>Aerodramus fuciphagus</i>	Aves	Apodiformes	Apodidae	LC
<i>Amaurornis phoenicurus</i>	Aves	Gruiformes	Rallidae	LC
<i>Anhinga melanogaster</i>	Aves	Pelecaniformes	Anhingidae	LC
<i>Ardea intermedia</i>	Aves	Pelecaniformes	Ardeidae	LC
<i>Ardea purpurea</i>	Aves	Pelecaniformes	Ardeidae	LC
<i>Bubulcus ibis</i>	Aves	Pelecaniformes	Ardeidae	LC
<i>Buceros rhinoceros</i>	Aves	Bucerotiformes	Bucerotidae	VU
<i>Centropus sinensis</i>	Aves	Cuculiformes	Centropidae	LC
<i>Dendrocygna arcuata</i>	Aves	Anseriformes	Anatidae	LC
<i>Excalfactoria chinensis</i>	Aves	Galliformes	Phasianidae	LC
<i>Gallinago stenura</i>	Aves	Charadriiformes	Scolopacidae	LC
<i>Gallirallus striatus</i>	Aves	Gruiformes	Rallidae	LC
<i>Geopelia striata</i>	Aves	Columbiformes	Columbidae	LC
<i>Gorsachius melanolophus</i>	Aves	Pelecaniformes	Ardeidae	LC
<i>Irena cyanogastra</i>	Aves	Passeriformes	Irenidae	NT
<i>Lophura ignita</i>	Aves	Galliformes	Phasianidae	VU
<i>Mixornis gularis</i>	Aves	Passeriformes	Timaliidae	LC
<i>Muscicapa latirostris</i>	Aves	Passeriformes	Muscicapidae	NA
<i>Myophonus caeruleus</i>	Aves	Passeriformes	Muscicapidae	LC
<i>Nycticorax nycticorax</i>	Aves	Pelecaniformes	Ardeidae	LC
<i>Phodilus badius</i>	Aves	Strigiformes	Tytonidae	LC
<i>Serilophus lunatus</i>	Aves	Passeriformes	Eurylaimidae	LC
<i>Todiramphus sanctus vagans</i>	Aves	Coraciiformes	Alcedinidae	LC
<i>Tricholestes criniger</i>	Aves	Passeriformes	Pycnonotidae	LC
<i>Aonyx cinereus</i>	Mammalia	Carnivora	Mustelidae	VU
<i>Callosciurus notatus</i>	Mammalia	Rodentia	Sciuridae	LC
<i>Callosciurus prevostii</i>	Mammalia	Rodentia	Sciuridae	LC
<i>Chironax sp.</i>	Mammalia	Chiroptera	Pteropodidae	LC
<i>Cynogale bennetti</i>	Mammalia	Carnivora	Viverridae	EN
<i>Cynopterus brachyotis</i>	Mammalia	Chiroptera	Pteropodidae	LC
<i>Cynopterus brachyotis</i>	Mammalia	Chiroptera	Pteropodidae	LC
<i>Echinosorex gymnura</i>	Mammalia	Eulipotyphla	Erinaceidae	LC
<i>Elephas maximus</i>	Mammalia	Proboscidea	Elephantidae	EN
<i>Galeopterus variegatus</i>	Mammalia	Dermoptera	Cynocephalidae	LC

<i>Hipposideros galeritus</i>	Mammalia	Chiroptera	Hipposideridae	LC
<i>Hylobates funereus</i>	Mammalia	Primates	Hylobatidae	EN
<i>Kerivoula pellucida</i>	Mammalia	Chiroptera	Vespertilionidae	NT
<i>Kerivoula sp.</i>	Mammalia	Chiroptera	Vespertilionidae	no
<i>Lenothrix canus</i>	Mammalia	Rodentia	Muridae	LC
<i>Lenothrix sp.</i>	Mammalia	Rodentia	Muridae	LC
<i>Lutra sumatrana</i>	Mammalia	Carnivora	Mustelidae	EN
<i>Lutrogale perspicillata</i>	Mammalia	Carnivora	Mustelidae	VU
<i>Macaca fascicularis</i>	Mammalia	Primates	Cercopithecidae	EN
<i>Macaca nemestrina</i>	Mammalia	Primates	Cercopithecidae	EN
<i>Manis culionensis</i>	Mammalia	Pholidota	Manidae	CR
<i>Manis javanica</i>	Mammalia	Pholidota	Manidae	CR
<i>Maxomys whiteheadi</i>	Mammalia	Rodentia	Muridae	VU
<i>Miniopterus fuliginosus</i>	Mammalia	Chiroptera	Vespertilionidae	NA
<i>Murina suilla</i>	Mammalia	Chiroptera	Vespertilionidae	LC
<i>Myotis horsfieldii</i>	Mammalia	Chiroptera	Vespertilionidae	LC
<i>Nasalis larvatus</i>	Mammalia	Primates	Cercopithecidae	EN
<i>Niviventer cremoriventer</i>	Mammalia	Rodentia	Muridae	LC
<i>Nycticebus menagensis</i>	Mammalia	Primates	Lorisidae	VU
<i>Paradoxurus jerdoni</i>	Mammalia	Carnivora	Viverridae	LC
<i>Paradoxurus hermaphroditus</i>	Mammalia	Carnivora	Viverridae	LC
<i>Penthetor lucasi</i>	Mammalia	Chiroptera	Pteropodidae	LC
<i>Pongo pygmaeus</i>	Mammalia	Primates	Hominidae	CR
<i>Potamochoerus</i>	Mammalia	Mammalia	Suidae	no
<i>Presbytis comata comata</i>	Mammalia	Primates	Cercopithecidae	EN
<i>Prionailurus bengalensis</i>	Mammalia	Carnivora	Felidae	LC
<i>Pteropus seychellensis comorensis</i>	Mammalia	Chiroptera	Pteropodidae	LC
<i>Pteropus sp.</i>	Mammalia	Chiroptera	Pteropodidae	EN
<i>Ptilocercus lowii</i>	Mammalia	Scandentia	Tupaiaidae	LC
<i>Rattus exulans</i>	Mammalia	Rodentia	Muridae	LC
<i>Rattus tanezumi</i>	Mammalia	Rodentia	Muridae	LC
<i>Rattus tiomanicus</i>	Mammalia	Rodentia	Muridae	LC
<i>Rhinolophus affinis</i>	Mammalia	Chiroptera	Rhinolophidae	LC
<i>Rhinolophus sp.</i>	Mammalia	Chiroptera	Rhinolophidae	no
<i>Sundamys muelleri</i>	Mammalia	Rodentia	Muridae	LC
<i>Trachypithecus cristatus</i>	Mammalia	Primates	Cercopithecidae	VU
<i>Trichys fasciculata</i>	Mammalia	Rodentia	Hystricidae	LC
<i>Tupaia longipes</i>	Mammalia	Scandentia	Tupaiaidae	LC
<i>Tupaia tana</i>	Mammalia	Scandentia	Tupaiaidae	LC
<i>Cuora amboinensis</i>	Reptilia	Testudines	Geoemydidae	EN
<i>Cyclemys sp.</i>	Reptilia	Testudines	Geoemydidae	NT
<i>Notochelys platynota</i>	Reptilia	Testudines	Geoemydidae	VU
<i>Varanus salvator</i>	Reptilia	Squamata	Varanidae	LC

Figure 31 : List of species

List of sites with location

SiteID	NameSite	X	Y	Water	Habitat
C1	CorridorB1P1	3129783	2015994	stagnant	Corridor
C2	CorridorB1P4A	3128857	2016490	stagnant	Corridor
C3	CorridorB1P5	3129161	2017534	stagnant	Corridor
C4	CorridorB1P6	3130258	2018964	stagnant	Corridor
C5	PlantationB2P5	3129149	2017512	stagnant	Corridor
E1	Estuary	3235002	2035119	flow	Forest
F1	ForestB3P3	3130398	2024063	stagnant	Forest
F10	RIVER MANAGBOL	3167009	2060862	flow	Forest
F11	RIVER TO DANAU PITAS ABAY	3168067	2063249	flow	Forest
F12	River ManagBol	3124393	2005851	flow	Forest
F2	ForestB3P3	3130604	2024069	stagnant	Forest
F3	ForestB3P5	3129214	2014627	stagnant	Forest
F4	Forest - KeruakBehind Bam's house	3129973	2010548	stagnant	Forest
F5	DANAU BACK	3169313	2065450	stagnant	Forest
F6	DANAU PITAS GAUCHE	3170648	2064695	stagnant	Forest
F7	OXBOW LAKE ENTREE KILINANAP	3118947	1996715	stagnant	Forest
F8	OXBOW LAKE FOND KILINANAP	3120254	1999203	stagnant	Forest
F9	OXBOW LAKE riv KILINANAP	3120549	1997631	flow	Forest
K1	Kinabatangan river	3164286	2065645	flow	Forest
K2	Kinabatangan river	3144434	2034518	flow	Mixte
K2	Kinabatangan river	3144434	2034518	flow	Mixte
K3	Kinabatangan river	3130848	2015798	flow	Forest
K3	Kinabatangan river	3130848	2015798	flow	Forest
K4	Kinabatangan river	3088307	1986732	flow	Mixte
K4	Kinabatangan river	3088307	1986732	flow	Mixte
K5	Kinabatangan river	3051751	1972233	flow	Forest
K5	Kinabatangan river	3051751	1972233	flow	Forest
K6	Kinabatangan river	2996018	1980909	flow	Forest
K6	Kinabatangan river	2996018	1980909	flow	Forest
K7	Kinabatangan river	2968478	1994835	flow	Forest
K7	Kinabatangan river	2968478	1994835	flow	Forest
K8	Kinabatangan river	2931465	1974513	flow	Mixte
K8	Kinabatangan river	2931465	1974513	flow	Mixte
M1	Mangrove	3216732	2016049	flow	Forest
O1	Oxbow lake	3170554	2062676	flow	Forest
O1	Oxbow lake	3170554	2062676	flow	Forest
O2	Oxbow lake	3117977	1998266	flow	Forest
O5	Oxbow lake	3011834	1972526	flow	Forest
P1	AgroforestryBlock A - Forest Native Species	3119863	2045328	stagnant	Plantation
P10	PlantationRiparian MOPP	3122942	2055992	stagnant	Plantation
P11	Agroforestry	3118118	2045070	stagnant	Plantation
P12	Agroforestry Block C	3121215	2044933	stagnant	Plantation

P13	Bridge Jambatan Orchid	3034436	2045814	stagnant	Plantation
P14	Hasbollah DUCK	3119896	2044780	stagnant	Plantation
P15	Small river M B95K	3119651	2051770	flow	Plantation
P2	AgroforestryBlock B - Mixed Native Species	3118411	2045003	stagnant	Plantation
P3	AgroforestryBlock C - Forest Island	3120594	2045384	stagnant	Plantation
P4	PlantationB2P10	3128416	2024751	stagnant	Plantation
P5	PlantationB2P6	3127323	2021854	stagnant	Plantation
P6	PlantationB2P8	3127324	2022949	stagnant	Plantation
P7	PlantationOil Palm MOPP	3120829	2053734	stagnant	Plantation
P8	PlantationOil Palm MOPP	3119826	2053707	stagnant	Plantation
P9	PlantationRiparian MOPP	3122316	2055566	stagnant	Plantation
PO1	Little river	3122697	2055135	flow	Plantation
T1	Little river	3154446	2073646	flow	Forest
T10	Little river	3128553	2015822	flow	Corridor
T11	Little river	3180368	2065346	flow	Forest
T11	Little river	3180368	2065346	flow	Forest
T2	Little river	3116857	1989774	flow	Forest
T2	Little river	3116857	1989774	flow	Forest
T3	Little river	2998786	1967778	flow	Forest
T4	Little river	2991022	2001177	flow	Plantation
T5	Little river	2956375	2002458	flow	Plantation
T6	Little river	2929870	1981208	flow	Forest
T6	Little river	2929870	1981208	flow	Forest
T7	Little river	3018452	1967186	flow	Forest
T8	Little river	3054923	1970132	flow	Forest
T9	Little river	3146876	2022311	flow	Plantation

Figure 32 : List of sites and locations

SiteID	Spygen	RICHESSSE SPECIFIQUE	Mammalia & Fish	MAMMALIA	REPTILIA	AVES	FISH	AMPHIBIAN
C1	SPY214361	19	12	7	1	1	5	5
C2	SPY214362	6	4	1	0	0	3	2
C3	SPY214366	8	4	4	0	2	0	2
C4	SPY214365	6	4	2	0	0	2	2
C5	SPY214364	11	7	2	1	2	5	1
E1	SPY193037	11	11	0	0	0	11	0
F1	SPY214369	22	17	17	0	2	0	3
F10	SPY223474	6	3	1	0	2	2	1
F11	SPY223469	6	6	3	0	0	3	0
F12	SPY214374	15	13	8	0	0	5	2
F2	SPY214370	16	11	8	1	2	3	2
F3	SPY214363	18	9	5	1	4	4	4
F4	SPY214372	7	4	4	1	1	0	1
F5	SPY223468	3	3	1	0	0	2	0
F6	SPY223466	2	2	0	0	0	2	0
F7	SPY223473	11	10	2	0	1	8	0
F8	SPY223467	7	6	1	0	1	5	0
F9	SPY223471	10	9	2	0	0	7	1
K1	SPY193021	4	4	1	0	0	3	0
K2	SPY193019	0	0	0	0	0	0	0
K2	SPY193027	2	2	1	0	0	1	0
K3	SPY193020	2	2	1	0	0	1	0
K3	SPY193028	0	0	0	0	0	0	0
K4	SPY193031	2	2	1	0	0	1	0
K4	SPY193048	3	3	1	0	0	2	0
K5	SPY193023	2	2	0	0	0	2	0
K5	SPY193040	5	5	2	0	0	3	0
K6	SPY193036	6	6	2	0	0	4	0
K6	SPY193039	6	6	3	0	0	3	0
K7	SPY193050	6	6	4	0	0	2	0
K7	SPY193054	7	7	3	0	0	4	0
K8	SPY193024	6	6	3	0	0	3	0
K8	SPY193032	9	9	7	0	0	2	0
M1	SPY193042	48	48	4	0	0	44	0
O1	SPY193038	4	4	1	0	0	3	0
O1	SPY193044	4	4	1	0	0	3	0
O2	SPY193043	3	3	1	0	0	2	0
O5	SPY193033	7	7	3	0	0	4	0
P1	SPY214375	7	3	1	0	3	2	1

P10	SPY223461	10	7	6	0	2	1	1
P11	SPY223472	7	4	1	0	2	3	1
P12	SPY223470	8	4	1	1	2	3	1
P13	SPY223464	8	6	3	0	1	3	1
P14	SPY223463	7	6	2	0	0	4	1
P15	SPY223465	21	15	9	1	4	6	1
P2	SPY214373	6	3	1	0	2	2	1
P3	SPY223462	10	5	2	0	4	3	1
P4	SPY214371	11	4	2	1	4	2	2
P5	SPY214367	9	5	3	1	3	2	0
P6	SPY214368	9	3	1	1	2	2	3
P7	SPY214376	8	5	3	0	1	2	2
P8	SPY223460	17	11	8	1	4	3	1
P9	SPY223459	17	7	5	1	6	2	3
PO1	SPY193030	1	1	0	0	0	1	0
T1	SPY193041	8	8	3	0	0	5	0
T10	SPY193025	4	4	2	0	0	2	0
T11	SPY193022	1	1	0	0	0	1	0
T11	SPY193029	1	1	0	0	0	1	0
T2	SPY193053	14	14	5	0	0	9	0
T2	SPY193055	11	11	4	0	0	7	0
T3	SPY193045	15	15	6	0	0	9	0
T4	SPY193052	10	10	2	0	0	8	0
T5	SPY193051	16	16	7	0	0	9	0
T6	SPY193034	13	13	6	0	0	7	0
T6	SPY193046	22	22	13	0	0	9	0
T7	SPY193047	8	8	2	0	0	6	0
T8	SPY193049	16	16	7	0	0	9	0
T9	SPY193026	17	17	7	0	0	10	0

Figure 33 : Biodiversity per sites